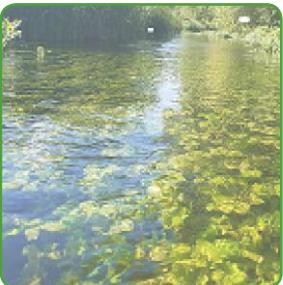
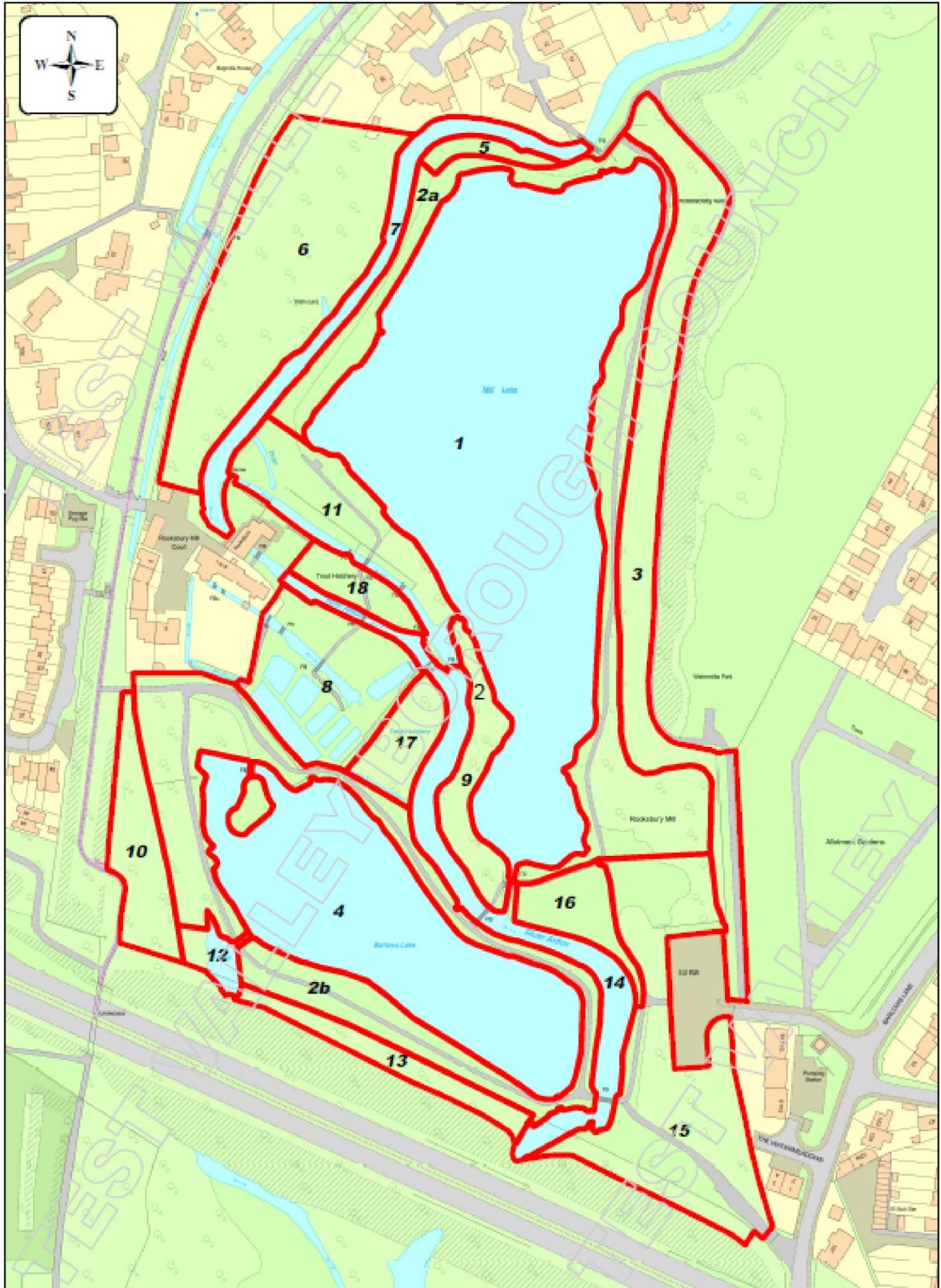


APPENDIX I

Compartment Map Descriptions Work Schedule Task based Risk Assessment





Scale: NTS @ A4 P
Date: 17 January 2024

**Map Showing Rooksbury Mill Local
Nature Reserve Compartment Boundaries**

Test Valley
Borough Council

Compartment 1	<i>Mill Lake</i>	<ul style="list-style-type: none"> • Cut banks yearly to end of the season and remove arisings. Cut on circular rotation - 1 third of bank per year (in accordance to EA regulations on flood prevention) • Areas between lakes become 'quiet' areas with no public access • Where possible, soften edges of lakes by decreasing gradient. • Cut aquatic weed three times yearly in April, June, August or as required and remove from site. • Manage trees around the lake to reduce wind shadow creating turbulent water to assist in the breakdown of ammonia and improve oxygen conditions. (See Appendix II - H) • Install Barley straw into the lake twice a year in January and June
Compartment 2a	<i>Improved pasture</i>	<ul style="list-style-type: none"> • Leave gap between paths and lakes • Cut grass to different swards heights. Mow and remove arisings. Where no formal path, cut grass as needed to maintain public access. Divide area into 3 parallel zones. Cut central area adjacent to paths twice yearly (first cut in March, second in October). Cut second zone (2-3 metres) every two years. Cut third zone on a four-year rotation • Control spread of Japanese knotweed using Roundup Biactive in compliance with EA regulations • Control ragwort through hand pulling and removal from site
Compartment 2b	<i>Improved pasture</i>	<ul style="list-style-type: none"> • Manage as in compartment 2a
Compartment 3	<i>Scrub boundary</i>	<ul style="list-style-type: none"> • Dependant on desire lines and entrances for pedestrians. • Coppice/pollard species when necessary • Gap up where possible using native species of local province
Compartment 4	<i>Barlows Lake</i>	<ul style="list-style-type: none"> • Manage banks as in compartment 1. • Increase light into wetland by promotory in Barlows Lake
Compartment 5	<i>Riparian planting</i>	<ul style="list-style-type: none"> • Coppice planted willow 1 in 3 annually from 2018 to maintain low level screening • Coppice planted alder from 2019 2 times annually to maintain low level screening
Compartment 6	<i>Riparian woodland</i>	<ul style="list-style-type: none"> • Remove trees if seen as a danger to the public • Leave as much dead wood as possible • Maintain and enhance emergent vegetation along bank • Remove trees from riverbank to increase light to river system, though leave a small number as bird perches
Compartment 7	<i>River Anton</i>	<ul style="list-style-type: none"> • Coppice overhanging trees • Encourage emergent vegetation by leaving a 2 meter buffer zone adjacent to the banks of the river

Compartment 8	<i>Meadows and stew ponds</i>	<ul style="list-style-type: none"> • Maintain meadow as in compartment 2 • Non-intervention to allow natural succession
Compartment 9	<i>Scrub</i>	<ul style="list-style-type: none"> • Non intervention • Restrict public and maintain no public access • Control breach in lake bank (overflow)
Compartment 10	<i>Poplar plantation</i>	<ul style="list-style-type: none"> • Phase out poplar following establishment of understory planting • Remove tree guards on planting when necessary • Coppice hazel on rotation from 2019
Compartment 11	<i>Rank fen</i>	<ul style="list-style-type: none"> • Maintain as fen. Cut on rotation one side each year • Thin trees as necessary to favour field layer
Compartment 12	<i>Rank fen</i>	<ul style="list-style-type: none"> • Thin trees around pond to increase light
Compartment 13	<i>Hedgerow</i>	<ul style="list-style-type: none"> • Maintain as hedgerow • Gap up where necessary using native trees of local province • Coppice to improve quality to allow for hedge to be laid in future
Compartment 14	<i>River Anton</i>	<ul style="list-style-type: none"> • Manage as in compartment 7 • Cut grass to different swards heights. Mow and remove arisings. Where no formal path, cut grass as needed to maintain public access. Divide area into 3 parallel zones. Cut central area adjacent to paths twice yearly (first cut in March, second in October). Cut second zone (2-3 metres) every two years. Cut third zone on a four-year rotation
Compartment 15	<i>Unimproved pasture</i>	<ul style="list-style-type: none"> • Cut grass twice yearly and remove arisings as in compartment 2 • Continue to remove scrub to favour nationally important ground flora.
Compartment 16	<i>Orchard</i>	<ul style="list-style-type: none"> • Practice regulated pruning on a 2 year rotation to maintain healthy trees and enhance the area for wildlife. Pruning should be conducted between November and the end of March, after the leaves have dropped and fruit has been produced. • Cut grass twice yearly removing arisings
Compartment 17	<i>Scrubland</i>	<ul style="list-style-type: none"> • Non-intervention to allow natural succession
Compartment 18	<i>Rough grassland</i>	<ul style="list-style-type: none"> • Control ragwort through hand pulling and removal from site • Cut and rake grass by hand and remove arising where possible • Control extent of bramble to avoid encroachment

Work Schedule 2024-2034

Objective	Prescription	Compartment	Year										
			2024	2025	2026	2027	2028	2029	2030	2031	2032	2033	2034
Maintain and increase habitat for water voles	Cut one third of the bank once per year (in accordance with EA regulations on flood prevention)	1,4,7,9,14	*	*	*	*	*	*	*	*	*	*	
Control Invasive ragwort	Pull by hand and remove from site	Whole Site	*	*	*	*	*	*	*	*	*	*	*
Make safe any dangerous trees	Remove unsafe trees from site. Leave deadwood on site if possible	Whole Site	*	*	*	*	*	*	*	*	*	*	*
Conduct surveys where necessary	Undertake surveys at appropriate time of year e.g. water vole, otter, bat, birds, terrestrial and aquatic invertebrates, higher and lower plant and reptile.												
	Bats	Whole Site	*			*	*			*			
	Water vole/otter	Whole Site	*		*					*			
	Birds	Whole Site		*	*								
	Terrestrial invertebrates	Whole Site		*	*			*					
	Aquatic invertebrates	Whole Site	*	*	*	*	*	*	*	*	*	*	*
	Higher and lower plants	Whole Site	*	*	*			*	*				
	Visitor	Whole Site			*				*				

Work Schedule 2024-2034													
Objective	Prescription	Compartment	Year										
			2024	2025	2026	2027	2028	2029	2030	2031	2032	2033	2034
	Butterflies	Whole Site	*	*	*	*	*	*	*	*	*	*	*
Monitor fishing within Mill Lake	Continue to bailiff fishing on Mill Lake	1	*	*	*	*	*	*	*	*	*	*	*
	Annual feedback questionnaire, including species list	1	*	*	*	*	*	*	*	*	*	*	*
Cut grass and remove arisings	Cut to differing sward heights twice yearly (first cut in March and second in October)	2a,2b,15,16	*	*	*	*	*	*	*	*	*	*	*
Keep footpaths accessible	Where no formal footpath exists cut grass twice yearly in March and October or as needed	2a,2b,8	*	*	*	*	*	*	*	*	*	*	*
Cut weed within Mill Lake to aid fishing	Cut and remove of site aquatic weed 3 times yearly in April June and September or as required	1	*	*	*	*	*	*	*	*	*	*	*
Maintain swims for fishing	Cut swims hedges and paths	1	*	*	*	*	*	*	*	*	*	*	*
	Maintain fishing platforms and replace where necessary	1	*	*	*	*	*	*	*	*	*	*	*
	Install new platforms on remaining swims (swims 1,3,5, 6, 8, 10, 11,12) one yearly	1	*	*	*	*							
Coppice/pollard	Coppice/pollard species where necessary to maintain healthy trees	6,7,14		*		*		*		*		*	

Work Schedule 2024-2034

Work Schedule 2024-2034



Risk Assessment

(Ref: RA/xx/xx)

To be used in conjunction with the following:

- Insert titles of other relevant risk assessments or policies

Activity:	General tasks	Location:	<i>Local Nature Reserves/ TVBC green space</i>
People at risk:	<i>Volunteers Staff Contractors</i>	Date:	<i>Jan 2026</i>
Completed by:	Daisy Kennard	Reviewed by:	<i>Charlotte Rimmer</i>

Location: Local Nature Reserves/ TVBC green space	Department: C&L
Description of Task: General tasks	Additional assessments required: No
Staff at Risk: Employee/ Member of Public/ Contractor/Volunteer	Risk Rating: <big>LOW</big>
Date of Assessment: 13.01.2026	Annually or as required.

No	Component of Task	Hazard	Nos. at Risk	Consequence	Existing Control Measures	Residual Risk Rating		
						L	S	Risk
1	Contact with water/surrounding land	Contact with biological diseases including Leptospirosis & Weils disease, Hep A, B, and C. Tetanus, Lyme disease, Avian influenza	1<	Serious illness, possible fatality	<ul style="list-style-type: none"> 1) Ticks - Keep arms and legs covered when walking in woodland/grassland. Check for ticks regularly and remove immediately if found. Light coloured clothing makes ticks easier to spot. Please see enclosed leaflet for details. 2) Cover wounds with a waterproof dressing. If a new wound occurs seek medical attention. 3) Clean hands with antibacterial hand wash or wipes before consuming food, drinking or smoking. 4) If symptoms develop seek medical attention immediately. 5) Do not touch needles. Do not touch dead animals or animal faeces. Refer to up-to-date DEFRA guidance on avian influenza. 	2	4	Med
2	Bending and kneeling in long vegetation	Contact with refuse, litter broken glass – general injury.	1	Minor injury/infection	<ul style="list-style-type: none"> 1) Inspect area for sharps/broken glass before starting an activity. 2) Clear any rubbish that is safe to do so or use alternative location. 3) Remind visitors of control measures. 4) Refer medical waste/syringes to group leader. Sharps box and 			

					PPE to be used by trained personnel only.			
3	Working close to deep/fast moving water	Falling in water	1	Minor injuries, exposure to cold/ hypothermia, risk of fatality from drowning	<ol style="list-style-type: none"> 1) Children under 16 must be accompanied by a responsible adult. 2) Assess suitability of site before entering the water. 3) Entry based on depth, flow, water quality, substrate type (gravel, silt etc.), bank condition and previous weather conditions. 4) Remind of control measures. 5) Supervising officer on water's edge should always carry a throwline. 6) Staff should have appropriate, up to date water safety training. 	1	3	Low
4	Working outside in cold/heat/wind/rain	Exposure to extreme elements- risk of hyperthermia or hypothermia, heatstroke, high winds/danger of falling objects	1<	Major illness/injury	<ol style="list-style-type: none"> 1) Adjust programme to avoid extremes. 2) Wear appropriate clothing, waterproofs, hat, sunscreen etc. 3) Take regular breaks and refreshments. 4) Have water available take regular breaks. 5) Remind of control measures. 6) Follow met office guidelines regarding weather warnings. 7) Do not run school visits during extreme temperatures 30 degrees + 	2	2	Low

5	Walking on non-surfaced routes	Slips, trips and falls	1<	Minor injuries such as twisted ankles, cuts and bruising	<ol style="list-style-type: none"> 1) Beware of potential uneven ground, rabbit holes etc. 2) Wear appropriate footwear with ankle support and good grip. 3) Inspect site before commencement of visit. 4) Identify areas of greatest risk and avoid. 5) Remind of control measures. 	4	1	Low
6	Walking/kneeling in areas accessible to Dogs	Contact with dog faeces/Toxocara	1	Serious illness/ infection (loss of sight)	<ol style="list-style-type: none"> 1) Be vigilant whilst walking around sites. 2) If contact with dog faeces occurs wash hands and other areas of contact thoroughly. 3) Do not touch face. 	1	3	Low
7	Interaction with members of the public	Violence and aggression	1<	Verbal/physical aggression	<ol style="list-style-type: none"> 1) Awareness of potential for aggressive or difficult members of the public. 2) Withdraw rather than face conflict. 3) Avoid lone working where possible. 4) Have functioning mobile phone available. 5) Use Orbis app where appropriate. 	1	3	Low
8	Contact with animals living on or visiting the site	Bites, scratches, Avian Influenza	1	Injury & infection from bites/ scratches	<ol style="list-style-type: none"> 1) Avoid contact with loose dogs. 2) Seek medical attention if bitten or scratched and report to police/dog warden and event organiser. 3) Refer to up-to-date DEFRA guidance on avian influenza 4) Up to date Tetanus vaccination. 5) Remind of controls. 	1	3	Low

					6) Do not touch dead animals or animal faeces			
9	Contact with biting/stinging insects	Allergic reaction, irritation	1<	Injury/infection/allergic reaction	<ol style="list-style-type: none"> 1) Inspect area prior to commencement of visit. 2) Do not walk near bee, wasp or hornet nests. 3) Be aware of signs and symptoms of anaphylaxis. 4) Phone emergency services immediately if symptoms of anaphylaxis or if sting victim known to be allergic. 5) Those with allergies should always have their medication available. 6) Clean sting/bite site immediately. 7) Alert group leader to any relevant allergies prior to commencement of visit. 	2	2	Low
10	Contact with Trees, shrubs and irritant/dangerous plants	allergic reaction	1	Injury/ infection/allergic reaction	<ol style="list-style-type: none"> 1) Awareness of risk. 2) Carry out site inspection before commencement of visit. 3) Identifying hazardous plants and avoid. 4) Wear long sleeves and trousers where possible. 5) Wash hands after contact and before eating or drinking or smoking 	2	2	Low

11	Working with tools to include - Bow saws/Loppers/Litter pickers/Secateurs/Rakes/Wheelbarrows/iron bar/driveall	Using sharp/heavy tools	1	Cuts, scratches, head injuries	<p>1) wear appropriate PPE (gloves/eye protection/hard hat/bump cap for iron bar and driveall use/steel toe boots) to protect from injury from inappropriate usage.</p> <p>2) Do not wear a glove on the hand holding the saw when felling trees.</p> <p>3) Do not hang saws/equipment from branches.</p> <p>4) Count tools back in after finishing.</p>	2	2	Low
12	Working with large/heavy/ multiple objects	Heavy objects, Manual handling	1	Injuries caused by manual handling	<p>1) Do not move or lift objects that are too heavy.</p> <p>2) Two handed lifts may be appropriate.</p> <p>3) Break down load into smaller manageable loads or seek assistance.</p> <p>4) Use good lifting technique. Ensure volunteers are comfortable in carrying out the task and have taken regular breaks</p> <p>5) Demonstrate correct lifting/manual handling technique</p>	2	2	Low

APPENDIX II

Species Records

A Fresh Water Invertebrates

B Floral Records

C Bird Records

D Bioblitz

E Moths

F Butterfly

G Water voles

H Fish Population Survey

I Invertebrate Survey

J Water Assessment and Management



A - Fresh Water Invertebrates

The data below has been gathered during school visits. A typical collecting day consists of four sessions. Six groups of children in each session collect from the water using nets with 1 mm mesh. These are sorted through in white trays and a selection of animals taken from the trays and put into smaller pots. Animals from all trays are used to compile a list for the day. Towards the end of the session, each group estimates the numbers of particular animal types in their tray. The list below shows the recorded species list for each year from 2014 - 2025.

Although there is considerable variation in the collecting effort and accuracy of each group, the number of repetitions does provide a degree of validity to the data, at least with respect to observing trends, and this is the main reason for gathering the data.

Animals recorded during the school visits

2020 - 2021 ALL SCHOOL VISITS CANCELLED DUE TO COVID

	2014	2015	2016	2017	2018	2019	2022	2023	2024	2025
Hydra										
Flatworm	*	*	*	*	*	*	*	*	*	*
Lumbriculus	*	*	*	*						*
Fish Leech – Piscicola	*	*		*	*	*		*	*	*
Leech – Theromyzon	*	*							*	*
Leech - Erpobdella	*	*		*						*
Amber Snail	*	*		*		*		*		
Snail - Bythinia	*	*	*	*	*	*		*		*
Bladder Snail	*		*							
Great Pond Snail	*	*	*	*	*	*		*	*	*
Ramshorn Snail	*	*	*	*	*	*		*	*	*
Orb Shell	*	*	*	*	*	*		*	*	
Waterflea - Daphnia		*	*	*	*	*		*	*	*
Waterflea - Cyclops	*	*	*	*	*	*				

Freshwater Shrimp	*	*	*	*	*	*		*	*	*
Hog-louse	*	*	*	*	*	*		*	*	*
Mayfly	*	*	*	*	*			*	*	
Hawker Dragonfly			*	*		*				*
Darter Dragonfly		*	*		*					
Damselfly	*	*	*	*	*	*		*	*	*
Water Measurer	*	*	*	*						*
Pond Skater	*	*	*	*	*	*	*	*	*	*
Water Cricket				*						
Water Scorpion	*	*	*	*	*	*	*	*	*	*
Water Stick insect	*	*	*	*	*					*
Saucer Bug	*					*				
Greater Water Boatman	*	*	*	*	*	*	*	*	*	*
Lesser Water Boatman	*	*	*	*	*	*	*	*	*	*
Caddis - stick case	*	*	*	*	*	*	*	*	*	*
Caddis - stone case	*	*	*	*	*	*	*	*	*	*
Caddis - green leaf case	*	*	*	*	*	*				
Alderfly	*	*								*
Giant Cranefly	*	*								
Phantom midge				*	*				*	*
Bloodworm	*	*	*	*	*	*		*	*	*
Biting midge		*	*	*	*					

B - Floral Records

Agrimony	Creeping bent
Amphibious bistort	Creeping buttercup
Annual Meadow Grass	Creeping cinquefoil
Apple	Creeping soft grass
Ash	Creeping thistle
Autumn hawkbit	Crested dogstail
Barren brome	Cut leaved crane's bill
Bee orchid	Cypress spurge
Bittersweet	Daffodil
Black horehound	Daisy
Blackthorn rowan	Dandelion
Blade Medic	Dark mullein
Blunt fruited water starwort	Devil's bit scabious
Bracken	Dogrose
Bramble	Dogwood
Branched bur reed	Elder
Broad leaved bamboo	Enchanters nightshade
Broad leaved dock	English Elm
Brooklime	Forget me knot
Buckthorn	Field horse tail
Buddleia	Fleabane
Bugle	Floating sweet grass
Bull rush	Flote / pilicate grass
Butterbur	Foxglove
Buttercup	Goosegrass
Cat's ear	Greater bird's foot trefoil
Charlock	Greater plantain
Chickweed	Great horse tail
Cleavers	Great mullein pot
Cocksfoot	Great willow herb
Columbine coltsfoot cowslip	Grey poplar
Common bent	Grey willow
Common birdsfoot trefoil	Ground elder
Common broom rape	Ground ivy
Common centaury	Groundsel
Common chickweed	Gypsy wort
Common club rush	Hairy sedge
Common comfrey	Hairy St John's wort
Common couch	Hairy tare
Common duck weed	Hard rush
Common figwort	Hart's tongue
Common horsetail	Hawthorn
Common ivy	Hazel
Common knapweed	Hedge bedstraw
Common mallow	Hedgerow
Common mouse-ear	Hedge woundwort
Common nettle	Hemlock hedge mustard
Common ragwort	Hemlock water dropwort
Common sorrel	Hemp agrimony
Common spotted orchard	Herb Robert
Common vetch	Himalayan cotoneaster
Compact rush	Himalayan giant bramble
Cow parsley	Hogweed
Crack willow	Honeysuckle
Crane's bill	Hyp radic

Floral Records

Ivy	Sheeps sorrel
Kidney vetch	Shepherd's purse
Knotgrass	Silver weed
Japanese knotweed	Smaller cat's tail
Lady fern	Smoothhawk's beard
Lady's bedstraw	Smooth sow-thistle
Large bindweed	Smooth-stalked meadow grass
Lesser burdock	Snowberry
Lesser spearwort	Soft rush
Lesser stitchwort	Southern marsh orchid
Lesser trefoil	Spear thistle
Lombardy poplar	Spiked sedge
Manes tail	Spindle
Marjoram	St John's wort (swuare stalked)
Marsh bedstraw	Stream water crowfort
Marsh foxtail	Sweet vernal grass
Marsh horsetail	Tall fescue
Marsh marigold	Tall oat
Marsh ragwort	Tormentil
Marsh thistle	Traveller's joy
Meadow buttercup	Tufted hairgrass
Meadow foxtail	Upright hedge parley
Meadow sweet	Wall barley
Monkey flower	Walnut
Mugwort	Water betony
Nipplewort	Water cress
Oval sedge	Water figwort
Parsnip	Water forget me knot
Peach leaved bellflower	Water mint
Perforate St John's wort	Water pepper
Pedunculate oak	Wavy bittercress
Perennial ox eye daisy	Weid
Perennial rye grass	Welted thistle
Pignut	White bryony
Pineapple weed	White clover
Prickly sow thistle	White dead nettle
Pyrimidal orchid	White water lily
Ragged robin	Wild angelica
Ratstail plantain	Wild carrot
Red bartsia	Wild cherry
Red campion	Wild marjoram
Red clover	Wild privet
Red fescus	Wild raspberry
Red-veined dock	Wild teasel
Reed sweet grass	Willow herb species
Remote sedge	Winter cress
Redshank	Wood avens
Reed canary grass	Wood dock
Ribwort plantain	Wood sedge
Rough hawkbit	Wych elm
Rough meadow grass	Yarrow
Russian comfrey	Yellow iris
Selfheal	Yellow rattle
Sharp flowered rush	Yorkshire fog

Barn Owl (2015)	Lapwing
Black Cap	Lesser Black Backed Gull
Blackbird	Lesser Redpoll
Black-headed Gull	Lesser Whitethroat
Blue Tit	Linnet
Brambling (2013)	Little Egret
Bullfinch	Little Grebe
Buzzard	Little Owl
Canada Goose	Long tailed Tit
Carriion Crow	Magpie
Cetti's Warbler (2014)	Mallard
Chaffinch	Mallard (domestic)
Chiffchaff	Marsh Tit
Coal Tit	Meadow Pipit
Collard Dove	Mistle Thrush
Common Crossbill (2016)	Moorhen
Common Gull	Mute swan
Common sandpiper (2013)	Nuthatch
Common Turn	Pheasant
Coot	Pied Wagtail (yarrellii)
Cormorant	Pied/White Wagtail
Cormorant (Continental) (2017)	Pochard
Cuckoo (2013)	Radd's Warbler (2013)
Dunnock	Raven
Feral Pigeon	Red Kite (2016)
Fieldfare	Red-legged Partridge
Firecrest (2016)	Redshank
Gadwall	Redwing
Garden Warbler (2014)	Reed Bunting
Goldcrest	Reed Warbler
Golden Plover	Ring-necked Duck
Goldfinch	Ring Ouzel
Great Black Backed Gull	Robin
Great Crested Grebe	Rook
Great Spotted Woodpecker	Sand Martin
Great Tit	Sedge Warbler
Great White Egret	Shovelor
Green Woodpecker	Siskin
Greenfinch	Skylark
Grey Heron	Song Thrush
Grey Wagtail	Sparrow Hawk
Greylag Goose	Spotted Flycatcher (2014)
Herring Gull	Starling
House Martin	Stock Dove
House Sparrow	Stonechat
Hybrid Aythya (2013)	Swallow
Hybrid Black x Mute Swan	Swift
Hybrid Duck	Tawny Owl
Jackdaw	Teal
Jay	Treecreeper
Kestrel	Tufted Duck
Kingfisher	Water Rail

Bird Records 2013 - 2019

Waxwing (2013)
Whitethroat (2013)
Wigeon
Willow Warbler (2014)
Woodcock
Wood Pigeon
Wren
Yellow browned warbler (2013)
Yellowhammer

Unidentified gull
Unidentified small gull

English name

Freshwater inverts

Banded damselfly nymph
Beetle larve
Biting midge larvae
Blue winged olive nymph
Bullhead fish
Cased caddisfly larvae
Caseless caddisfly nymph
Flatworm
Freshwater limpet
Freshwater shrimp
Greater water boatnab
Hairworm
Hoglouse
Mayfly nymph
Midge larvae
Pond skater
Pond snail
Ramshorn snail
Signal crayfish
Stick cased caddisfly
Stone cased caddisfly
Stonefly nymph
Wandering snail
Water mite
Water scorpion
Whirlgig beetle

Damselflies

Banded Damselfly
Common blue damselfly

Mammals

Otter tracks (on mink raft)

Butterflies

Common blue
Large white
Meadow brown
Painted lady
Red admiral
Small skipper
Small tortoise shell
Small white

Orchids

Bee orchid
Pyramid orchid

Bees

Common Carder Bee
Buff-tailed Bumblebee

Moths

Scarlett tiger moth

E - Moths identified at Rooksbury Mill LNR

These moths were attracted to two light sources set up just north of the car park by Alison Cross and Mike Wall of Butterfly Conservation (Hampshire). The light sources were run from 21.45 to 23.30. Weather was calm, warm but overcast. There was significant bat activity. The moths were shown to a large group of local residents who enjoyed this event that had been programmed by Hampshire and Isle of Wight Wildlife Trust (NW District) and advertised locally.

English name	Scientific name	Comment
	* <i>Pyrausta aurata</i>	Also seen in daylight
Shuttle-shaped Dart	<i>Agrotis puta</i> ssp. <i>puta</i>	
	<i>Lathronympha strigana</i>	
Mother of Pearl	* <i>Pleuroptya ruralis</i>	Also seen in daylight
	<i>Agapeta hamana</i>	
Brimstone	<i>Opisthograptis luteolata</i>	
Dwarf Cream Wave	<i>Idaea fuscovenosa</i>	
	<i>Dipleurina lacustrata</i>	
Pebble Hook-tip	<i>Drepana falcata</i> <i>falcata</i>	
Dun-bar	<i>Cosmia trapazina</i>	
Single-dotted Wave	<i>Idaea dimidiata</i>	
	<i>Paraswammerdamia nebulella</i>	
Yellow Shell	<i>Camptogramma bilineata</i> <i>bilineata</i>	Also seen in daylight
Flame Shoulder	<i>Ochropleura plecta</i>	
	<i>Trachycera advenella</i>	
Dingy Footman	<i>Eilema griseola</i>	
Round-winged Muslin	<i>Thumatha senex</i>	
Red Twin-spot Carpet	<i>Xanthorhoe spadicearia</i>	
	<i>Phyllonorycter emberizaepenella</i>	
The Flame	<i>Axylia putris</i>	
Dark-barred Twin-spot Carpet	<i>Xanthorhoe ferrugata</i>	
Common Carpet	<i>Epirrhoa alternata</i> <i>alternata</i>	Also seen in daylight
Small Magpie	* <i>Eurrhypara hortulata</i>	
	<i>Chrysoteuchia culmella</i>	
Early thorn	<i>Selenia dentaria</i>	
Ruby Tiger	<i>Phragmatobia fuliginosa</i>	
Cloaked minor	<i>Mesoligia furuncula</i>	
Large Yellow Underwing	<i>Noctua pronuba</i>	

English name	Scientific name	Comment
White satin	<i>Leucoma salicis</i>	
	<i>Batia unitella</i>	
Buff Ermine	<i>Spilosoma luteum</i>	
Setaceous Hebrew Character	<i>Xestia c-nigrum</i>	
	<i>Yponomeuta evonymella</i>	
Common Wave	<i>Cabera exanthemata</i>	
	<i>Bryotropha terella</i>	
	<i>Agonopterix heracliana</i>	
	<i>Borkhausenia fuscescens</i>	
Sallow Kitten	<i>Ypsolopha scabrella</i>	
	<i>Furcula furcula</i>	
Riband Wave	<i>Idaea aversat</i>	
Ringed China-mark	* <i>Parapoynx stratiotata</i>	
	<i>Blastobasis adustella</i>	
	<i>Euzophera pinguis</i>	
Smoky Wainscot	<i>Mythimna impura</i>	
	<i>Yponomeuta padella</i>	
	<i>Acleris aspersana</i>	
Yellow-tail	<i>Euproctis similis</i>	Also seen in daylight
	<i>Cnephacia sp.</i>	
Willow Beauty	<i>Peribatodes rhomvoidaria</i>	
V-Pug	<i>Chloroclystis v-ata</i>	
Clouded Border	<i>Lomaspilis marginata</i>	
Elephant Hawkwoth	<i>Deilephila elpenor</i>	Larvae found 22/10/2010

English Name	Scientific Name	Comment
<i>Anticlea derivata</i>	Streamer	
<i>Ectropis bistortata</i>	Engrailed	
<i>Gymnoscelis rufifasciata</i>	Double-striped pug	
<i>Orthosia gothica</i>	Hebrew character	
<i>Anthophila fabriciana</i>	Nettle-tap	
<i>Caloptilia syringella</i>		
<i>Micropterix calthella</i>		
<i>Acronicta rumicis</i>	Knot grass	
<i>Depressaria heraclei</i>	Parsnip moth	
<i>Parapoynx stratiotata</i>	Ringed china-mark	
<i>Callimorpha dominula</i>	Scarlet tiger moth	
<i>Cameraria ohridella</i>	Horse-chestnut leaf-miner	
<i>Celypha lacunana</i>		
<i>Chrysoteuchia culmella</i>		
<i>Dichrorampha sequana</i>		
<i>Timandra comae</i>	Blood-vein	
<i>Ennomos fuscantaria</i>	Dusky thorn	
<i>Hypena proboscidalis</i>	Snout	
<i>Morma maura</i>	Old lady	
<i>Noctua fimbriata</i>	Broad-bordered yellow underwing	
<i>Orgyia antiqua</i>	Vapourer	
<i>Xestia c-nigrum</i>	Setaceous hebrew character	
<i>Xestia xanthographa</i>	Square-spot rustic	

*marked species are Pyralid Moths

This list will be submitted for the National and Hampshire records by the moth experts mentioned above.

List provided by Alison Cross. Elephant Hawkmoth larvae note added by Mervyn Grist.

On this date some additional species of moths were recorded from the footpath underpass (of A303) just west of Rooksbury Mill. These were:

Black Arches	<i>Lymantria monacha</i>
Large twin-spot carpet	<i>Xanthorhoe quadrifasiata</i>

F - Butterfly Survey 2025

Rooksbury Mill Local Nature Reserve Butterfly Survey **Annual Report 2025**

A Big Thank You

First off, I would like to thank everybody who participated in this year's butterfly surveys. Test Valley Borough council are very grateful to have amazing volunteers who conduct surveys to give us such a wide range of results. Thank you all for collecting such valuable data.

Introduction

The Rooksbury Mill Local Nature Reserve Butterfly transect was set up in 2010 and during this time has had 29 species of butterfly recorded on site. This site has a variety of different habitats which benefit a wide range of species.

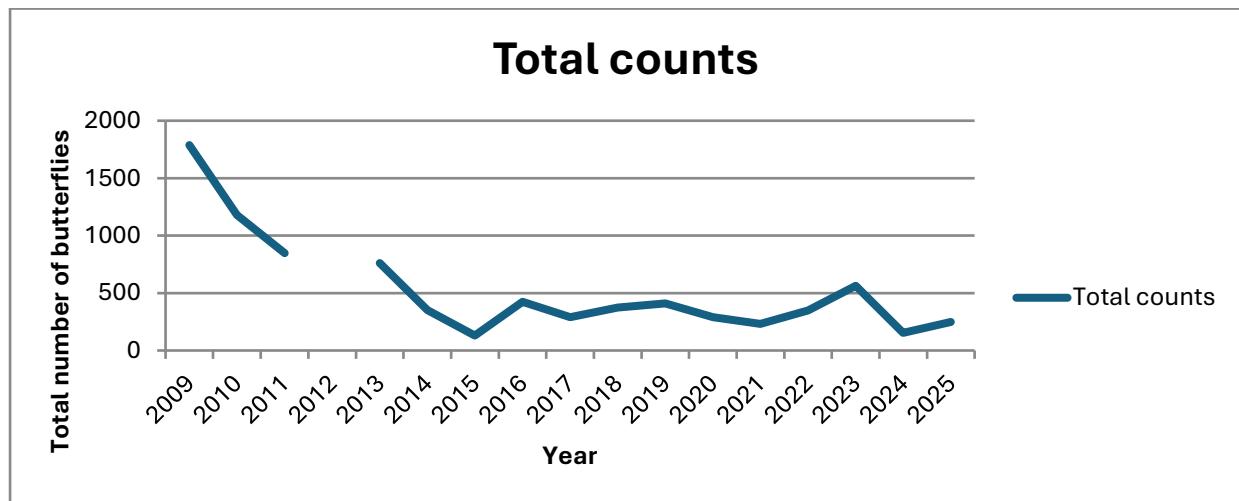
Results

This year, a total of 249 butterflies were recorded across 18 visits. An average of 13.83 butterflies were recorded per survey, a 53% increase compared with last year (17 visits).

This year, butterfly abundances peaked at 60 individuals on 27th June, which was earlier and higher than 2023. The highest counts from this peak were Small White (15) and Large white (14).

The most abundant species this year was the Small White, with 94 counted in total; an average of 5.2 individuals per survey. Last year the Large White was the most abundant species with 32 individuals recorded, with an average of 1.88 per survey. This year, Small White counts increased by 520% from 18 sightings to 94.

The second and third most abundant species were the Large White and Peacock, with averages 2.38 and 1.2 per survey respectively.



This year, 1 Purple Hairstreak was recorded at Rooksbury, the first recorded since records began in 2010. 5 Essex Skippers were recorded this year, creating a gap from previous sightings in 2022. Compared to last year's recordings, there were no sightings of Holly Blue or Marbled White.

As only 18 surveys were completed this year, the minimum of 24 surveys recommended by the UK Butterfly Monitoring Scheme to enable accurate conclusions to be drawn was not met.

A Full list of species from 2025 can be seen below with the six most common highlighted in **bold**:

Small Skipper	Purple Hairstreak	Peacock
Essex Skipper	Orange Tip	Comma
Brimstone	Common Blue	Speckled Wood
Large White	Red Admiral	Gatekeeper/Hedge Brown
Small White	Painted Lady	Meadow Brown
Green-veined White	Small Tortoiseshell	Ringlet

Conclusion

In conclusion, overall butterfly abundances at Rooksbury Mill Local Nature Reserve increased by approximately 61% this year compared with 2024. The Small White was the most abundant butterfly species recorded at the site this year, replacing the Large White whose numbers showed a slight increase of 34% from 2024-2025. A total of 18 species were recorded in 2025 compared to 17 in 2024; however, both years had fewer site visits than 2023, which had 20 species recorded for 21 site visits.

G - Watervole Survey – Map



Scale:
Date:
Drawn:
Dept:
Doc:

**Test Valley Borough
Council Mapping**

Test Valley
Borough Council

National Water Vole Monitoring Programme

recording forms

people's
trust for
endangered
species

Please ensure you have read the survey guidelines before doing your survey. Please complete the *Site information form* when you do your preliminary visit of your site. You will then need to complete the *NWVMP survey form* when you do your survey in May.

Site information form

1. Site number: Rooksbury Mill LNR

2. Habitat:

Upland

Lowland

Coastal

3. Waterway type:

river

stream/burn

lake/loch

reed bed

marsh

bog

pond

canal

reservoir

ditch/dyke

other

If other please specify:

4. Have water voles been reintroduced at this site?

Yes No Don't know

If yes, please specify when the reintroduction happened:

5. Total number of transects at site (usually one):1.....

6. Does mink control occur on this site?

Yes No Don't know

Mink surveys undertaken.

Your Transect details

7. Start grid reference (8 figure grid reference e.g. XX74736789) ...SU356447.....

8. End grid reference (8 figure grid reference e.g. XX74736789) ...SU360448.....

9. Grid references of each 100m mark along your transect (complete as many as is applicable to the length of your transect – you will not be asked to enter this data online):

100m	200m	300m	400m	500m	600m
SU357447	SU358448	SU358449	SU359449	SU360448	

National Water Vole Monitoring Programme survey form

Site Number:	Transact No. (if applicable): 2	Bank surveyed (N/S/E/W): Both
Start grid ref: SU356447	End grid ref: SU360448	Transect length: 500m
Survey date: 30/07/2017	Surveyor: Ron Davis and Kate Savage	

Water vole signs in each 100m section of the transect (complete as many as is applicable to the length of your transect)					
Field sign	0-100m	100-200m	200-300m	300-400m	400-500m
Number of trampled latrines (trodden flat on top)	4		1		500-600m
Number of untrampled latrines	3		4		
Please note the location of the <i>first</i> latrine that you encounter (Grid Reference or GPS)	SU359449	SU358449	SU358449	SU360447	
Please note the location of the <i>last</i> latrine that you encounter (Grid Reference or GPS)					
Water vole feeding signs					
Burrows/nests (approximate no.)	<input checked="" type="checkbox"/> None <input type="checkbox"/> 1-5 <input type="checkbox"/> 6-10 <input type="checkbox"/> More than 10	<input checked="" type="checkbox"/> None <input type="checkbox"/> 1-5 <input type="checkbox"/> 6-10 <input type="checkbox"/> More than 10	<input type="checkbox"/> None <input checked="" type="checkbox"/> 1-5 <input type="checkbox"/> 6-10 <input type="checkbox"/> More than 10	<input type="checkbox"/> None <input checked="" type="checkbox"/> 1-5 <input type="checkbox"/> 6-10 <input type="checkbox"/> More than 10	<input checked="" type="checkbox"/> None <input type="checkbox"/> 1-5 <input type="checkbox"/> 6-10 <input type="checkbox"/> More than 10
Sighting:	<input type="checkbox"/> No <input type="checkbox"/> Yes, if so how many:	<input type="checkbox"/> No <input type="checkbox"/> Yes, if so how many:	<input checked="" type="checkbox"/> No <input type="checkbox"/> Yes, if so how many:	<input checked="" type="checkbox"/> No <input type="checkbox"/> Yes, if so how many:	<input type="checkbox"/> No <input type="checkbox"/> Yes, if so how many:

Mink/otter signs along the whole transect (only record if you are <i>certain</i> of your identification and please take a photo including an object to indicate scale)	
Mink	Otter
<input type="checkbox"/> Scat	<input type="checkbox"/> Spraint
<input type="checkbox"/> Footprints	<input type="checkbox"/> Footprints
<input type="checkbox"/> Sighting	<input type="checkbox"/> Sighting
Mink raft present <input type="checkbox"/> Yes <input type="checkbox"/> No	
Any comments on mink control at the site/transect	

*Data should be submitted by the 31st October each year
Please enter all data online at www.ptes.org/watervoles*

Field sign record form (optional)

Please record the field sign detected (trampled/untrampled latrine, feeding station, burrow, sighting) and its GPS location			
Field sign	Location (Grid Reference or GPS)	Transect section: e.g. 1 st 100m, 3 rd 100m	Comments
e.g. trampled latrine	SN6447725383 or Lat: 51.910444 Long: -3.9716113	1 st 100m section	Fresh droppings

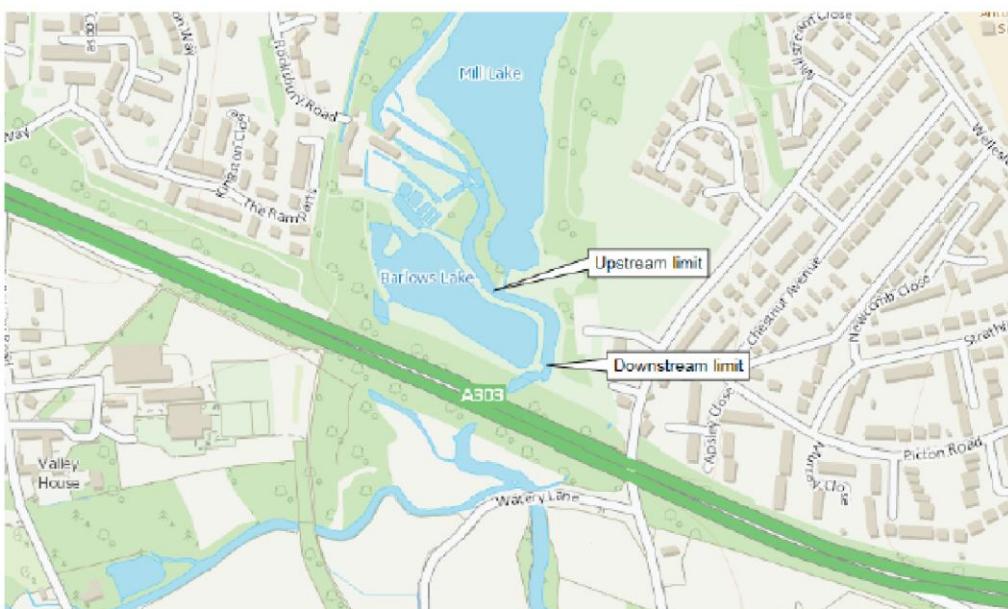
Fish population survey summary: Rooksbury Mill, River Anton

Background

A fish population survey was carried out on the River Anton at Rooksbury Mill, Andover on the 8th September, 2016. The survey was part of the six-yearly Salmon Action Plan survey programme, aimed at assessing the spatial distribution of juvenile salmon throughout the Test catchment.

Methods

A single electric fishing run was completed over an 85m reach between SU3564844305 and SU3561144378 (Map 1, below). A battery-powered, backpack electric fishing unit was used, with one anode. Two team members captured fish in dip nets and another carried an aerated holding tank. The average width of the survey reach was 5.27m. Captured fish were identified, measured and returned to the river.



Map 1: Survey location

Results

The survey catch consisted of 68 brown trout *Salmo trutta*, 92 bullhead *Cottus gobio* and 4 European eels *Anguilla anguilla*, as shown in the pie-chart below:

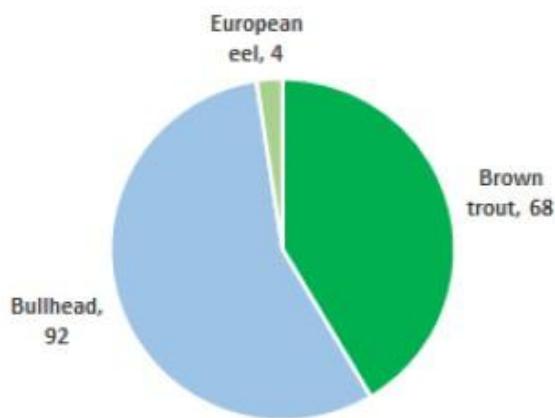


Figure 1: Survey catch

The lengths of the eels captured are given in figure 2, below:

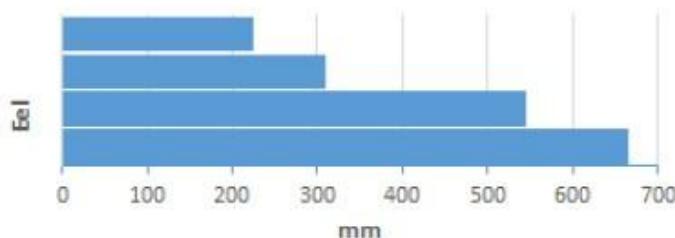


Figure 2: Eel lengths

Figure 3 is a length frequency histogram showing the numbers of trout in each size category.

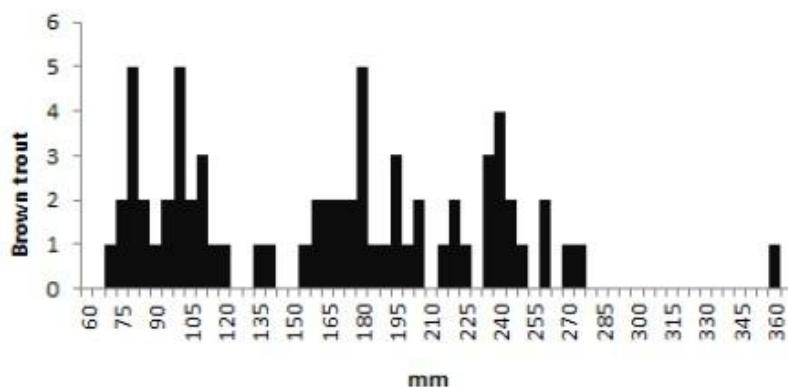


Figure 3: Brown trout length frequency (n=68)



Discussion

The total catch of brown trout in 2016 was lower (68) than when the survey was conducted in 2010 (129). However, the average width of the site was 9.6m in 2010 and a 100m reach was fished, whereas in 2016 the average width was 5.27m and an 85m reach was fished. The reduction in width was probably due to a combination of lower flow and development marginal vegetation. The reach length was reduced in order to align with a suitable upstream stop net position. This means that brown trout density in 2016 was 15.2 per 100m², slightly higher than in 2010 when it was 13.5 per 100m².

Figure 3 indicates fairly even numbers of juvenile, mid-aged and mature brown trout, which is a reflection of good habitat quality and complexity - in order for a reach to support these different life stages, it must meet their varied habitat requirements (i.e. pools, riffles, variable depth & substrate).

The capture of 92 bullhead indicates both good water quality and good substrate (riverbed) habitat. Bullhead are less affected by larger scale habitat features but require complex, stony substrate, ideally with woody debris and vegetated margins.

The eel catch was similar to 2010, when three eels were caught (of similar lengths to 2016). The European eel is critically endangered and abundance, especially in headwaters, has declined substantially in recent decades. The Rooksbury catches demonstrate that the site is accessible to eels migrating upstream from the sea and that it provides suitable habitat quality and food availability to support resident adult eels.

The 2010 survey also recorded nine grayling, one three-spined stickleback, one brook lamprey, one stone loach and three juvenile pike. The absence of grayling from the 2016 catch may reflect the general decline in this species that has been noted in recent years, probably as a result of high temperatures and low flows. The absence of the other species is probably a random feature of the results and not a cause for concern.

INVERTEBRATE SURVEY OF

ROOKSBURY MILL,

ANDOVER

NORTH HAMPSHIRE

NOVEMBER 2019

Dr. Jonty Denton FRES FLS CEcol MCIEEM

31 Thorn lane, Four Marks, Hants, GU34 5BX

Summary

A survey of terrestrial and aquatic invertebrates was carried out across the site in April-September 2019.

Survey date/s: 23rd May, 27th June, 1st, 11th July & 15th August & 10th September 2019

Species total: A total of 364 invertebrate taxa were identified of which 9 had conservation statuses. These are listed below;-

Species	Family	Order		Conservation status
<i>Theridiosoma gemmosum</i>	Theridiosomatidae	Araneae	Ray spider	NS
<i>Paracorymbia fulva</i>	Cerambycidae	Coleoptera	A longhorn beetle	RDB 3
<i>Agelastica alni</i>	Chrysomelidae	Coleoptera	Alder leaf beetle	DD;NR
<i>Plateumaris rustica</i>	Chrysomelidae	Coleoptera	A reed beetle	NS
<i>Drupenatus nasturtii</i>	Curculionidae	Coleoptera	A weevil	[Nb]
<i>Gymnetron veronicae</i>	Curculionidae	Coleoptera	A weevil	Nb
<i>Gymnetron villosulum</i>	Curculionidae	Coleoptera	A weevil	[Nb]
<i>Elodes elongata</i>	Scirtidae	Coleoptera	A marsh beetle	NS
<i>Aquarius paludum</i>	Gerridae	Hemiptera	Large Pondskater	NS

INTRODUCTION

The project brief was to provide baseline records for invertebrates across the site.

METHODOLOGY AND SITE VISITS

The main emphasis of the survey was to find as many rare and notable species as possible within the reviewed groups.

The site was visited specifically for invertebrate surveying on the following dates;- 23rd May, 1st, 11th & 31st July & 18th August 10th & 21st September 2019

Standard field techniques were employed to sample the invertebrate fauna across the site. These included sweeping vegetation with a wide mouthed sweep net, beating trees and bushes over a beating tray, and grubbing amongst tussocks and key host plant rosettes etc.

Because it is impracticable to survey all the potential invertebrates within any given site, only specific groups of species were examined during fieldwork. These groups are sufficiently well known as to allow meaningful comparisons to be made with other sites, both locally and nationally. They are also important as indicators of the quality of a site and the habitats present (see Brooks 1993).

Groups covered during the survey were:

- Mollusca (slugs and snails)
- Arachnida (spiders, harvestmen & pseudoscorpions)
- Isopoda (woodlice)
- Thysanura (bristletails)
- Ephemeroptera (mayflies)
- Odonata (dragonflies & damselflies)
- Plecoptera (stoneflies)
- Orthoptera (grasshoppers & crickets)
- Dictyoptera (cockroaches)
- Dermaptera (earwigs)
- Hemiptera-Heteroptera (true-bugs)
- Hemiptera-Homoptera (hoppers)
- Neuroptera (lace-wings)
- Mecoptera (scorpion-flies)
- Lepidoptera (butterflies & moths)
- Trichoptera (caddis flies)
- Diptera (true flies)
- Aculeate Hymenoptera (ants, bees & wasps)
- Coleoptera (beetles)

Measuring the quality of other invertebrate fauna

The invertebrate assemblages present at the site during the survey are assessed using Natural England's Invertebrate Species-habitat Information System (ISIS – 2010 version), as defined in Webb & Lott (2006) and Drake *et al.* (2007). Further developments for the programme are discussed in Lott (2008).

The system was developed for Common Standards Monitoring (CSM) on Sites of Special Scientific Interest (SSSI) but other applications are possible at a range of geographic scales. Lott (2008) describes the essence of ISIS as a database that can be used to recognise invertebrate assemblage types in species lists and evaluate their value for nature conservation.

RESULTS

A total of 364 species of invertebrate were recorded. A full species list with UK statuses is given in Appendix 1. Of these, twenty species have a conservation designation: These are summarised in Table 1. IUCN re-evaluated species have their IUCN criteria given followed by their current UK rarity status in brackets. Those species that have not yet been IUCN re-evaluated have their current statuses in square brackets. The definitions for these criteria are given in appendix 2.

Table 1. list of species with a conservation designation.

Species	Family	Order		Conservation status
<i>Theridiosoma gemmosum</i>	Theridiosomatidae	Araneae	Ray spider	NS
<i>Paracorymbia fulva</i>	Cerambycidae	Coleoptera	A longhorn beetle	RDB 3
<i>Agelastica alni</i>	Chrysomelidae	Coleoptera	Alder leaf beetle	DD;NR
<i>Plateumaris rustica</i>	Chrysomelidae	Coleoptera	A reed beetle	NS
<i>Drupenatus nasturtii</i>	Curculionidae	Coleoptera	A weevil	[Nb]
<i>Elodes elongata</i>	Scirtidae	Coleoptera	A marsh beetle	NS
<i>Aquarius paludum</i>	Gerridae	Hemiptera	Large Pondskater	NS

UKBAP / SPI (NERC S41) species:

Tyria jacobaeae - Cinnabar

SURVEY LIMITATIONS

The moth fauna is always under-represented when only diurnal surveys are employed. Light trapping surveys with 2-3 MV and actinic Robinson type traps would add hundreds of species of moth and additional night flying species (Ichneumonidae, Coleoptera etc.)

ECOLOGICAL ASSESSMENT

Rooksbury Mill continues to support a rich assemblage with excellent river margin habitats.

The excessive algal development in the main lake continues to be a blight on the reserve

REFERENCES

Brooks, S.J. 1993. Joint Committee for the Conservation of British Invertebrates: Guidelines for Invertebrate Surveys. *British Wildlife* 4(5) 283-287

Harvey, P.R., Nellist, D.R. & Telfer, M.G. (eds) 2002. *Provisional Atlas of British Spiders (Arachnida, Araneae) Volumes 1 & 2*. Huntingdon: BRC.

Hyman, P.S & Parsons, M.S. 1992. *A review of the scarce and threatened Coleoptera of Great Britain. Part 1*. JNCC, Peterborough.

Kirby, P. 1992. *A review of the scarce and threatened Hemiptera of Great Britain*. Peterborough, JNCC.

Merrit, P. 1990. *A review of the Nationally Notable Spiders of Great Britain*. Peterborough, NCC.

APPENDICES

Table 1. Species list 2019

Species	Family	Order	Conservation status
<i>Aceria erinea</i>	Eriophyidae	Prostigmata	common
<i>Acilius sulcatus</i>	Dytiscidae	Coleoptera	common
<i>Adalia bipunctata</i>	Coccinellidae	Coleoptera	common
<i>Agabus bipustulatus</i>	Dytiscidae	Coleoptera	common
<i>Agabus didymus</i>	Dytiscidae	Coleoptera	common
<i>Agabus nebulosus</i>	Dytiscidae	Coleoptera	common
<i>Agabus paludosus</i>	Dytiscidae	Coleoptera	common
<i>Agelastica alni</i>	Chrysomelidae	Coleoptera	DD;NR
<i>Aglais io</i>	Nymphalidae	Lepidoptera	common
<i>Aglais urticae</i>	Nymphalidae	Lepidoptera	common
<i>Agonum fuliginosum</i>	Carabidae	Coleoptera	common
<i>Agriphila geniculea</i>	Crambidae	Lepidoptera	common
<i>Alboglossiphonia heteroclita</i>	Glossiphoniidae	Rhynchobdellida	common
<i>Alianta incana</i>	Staphylinidae	Coleoptera	common
<i>Alnetoidea alneti</i>	Cicadellidae	Hemiptera	common
<i>Altica lythri</i>	Chrysomelidae	Coleoptera	common
<i>Amalorhynchus melanarius</i>	Curculionidae	Coleoptera	common
<i>Amaurobius fenestralis</i>	Amaurobiidae	Araneae	common
<i>Amblyteles armatorius</i>	Ichneumonidae	Hymenoptera	common
<i>Anacaena globulus</i>	Hydrophilidae	Coleoptera	common
<i>Anasimyia contracta</i>	Syrphidae	Diptera	common
<i>Anaspis fasciata</i>	Scaptiidae	Coleoptera	common
<i>Anaspis maculata</i>	Scaptiidae	Coleoptera	common
<i>Anaspis regimbarti</i>	Scaptiidae	Coleoptera	common
<i>Anax imperator</i>	Aeshnidae	Odonata	common
<i>Anisosticta novemdecimpunctata</i>	Coccinellidae	Coleoptera	common
<i>Anobium punctatum</i>	Anobiidae	Coleoptera	common
<i>Anthocharis cardamines</i>	Pieridae	Lepidoptera	common
<i>Anthocoris confusus</i>	Anthocoridae	Hemiptera	common
<i>Anthocoris limbatus</i>	Anthocoridae	Hemiptera	common
<i>Anthocoris nemoralis</i>	Anthocoridae	Hemiptera	common
<i>Anthonomus pedicularius</i>	Curculionidae	Coleoptera	common
<i>Anyphaena accentuata</i>	Anyphaenidae	Araneae	common
<i>Aphthona euphorbiae</i>	Chrysomelidae	Coleoptera	common
<i>Aphthona nonstriata</i>	Chrysomelidae	Coleoptera	common
<i>Apis mellifera</i>	Apidae	Hymenoptera	common
<i>Aquarius paludum</i>	Gerridae	Hemiptera	NS
<i>Araneus diadematus</i>	Araneidae	Araneae	common
<i>Araneus marmoreus</i> var. <i>pyramidalatus</i>	Araneidae	Araneae	local

<i>Archarius salicivorus</i>	Curculionidae	Coleoptera	common
<i>Arge cyanocrocea</i>	Argidae	Hymenoptera	common
<i>Argyresthia pygmaeella</i>	Argyresthiidae	Lepidoptera	common
<i>Asellus (Asellus) aquaticus</i>	Asellidae	Isopoda	common
<i>Athalia rosae</i>	Tenthredinidae	Hymenoptera	common
<i>Atrecus affinis</i>	Staphylinidae	Coleoptera	common
<i>Bibio marci</i>	Bibionidae	Diptera	common
<i>Bithynia tentaculata</i>	Bithyniidae	Littorinimorpha	common
<i>Blepharidopterus angulatus</i>	Miridae	Hemiptera	common
<i>Bombus hypnorum</i>	Apidae	Hymenoptera	common
<i>Bombus lapidarius</i>	Apidae	Hymenoptera	common
<i>Bombus lucorum</i>	Apidae	Hymenoptera	common
<i>Bombus pascuorum</i>	Apidae	Hymenoptera	common
<i>Bombus pratorum</i>	Apidae	Hymenoptera	common
<i>Byturus tomentosus</i>	Byturidae	Coleoptera	common
<i>Calameuta filiformis</i>	Cephidae	Hymenoptera	common
<i>Calameuta pallipes</i>	Cephidae	Hymenoptera	common
<i>Callimorpha dominula</i>	Erebidae	Lepidoptera	local
<i>Calopteryx splendens</i>	Calopterygidae	Odonata	common
<i>Calvia quatuordecimguttata</i>	Coccinellidae	Coleoptera	common
<i>Cantharis cryptica</i>	Cantharidae	Coleoptera	common
<i>Cantharis decipiens</i>	Cantharidae	Coleoptera	common
<i>Cantharis lateralis</i>	Cantharidae	Coleoptera	common
<i>Cantharis nigricans</i>	Cantharidae	Coleoptera	common
<i>Cantharis rustica</i>	Cantharidae	Coleoptera	common
<i>Capsus ater</i>	Miridae	Hemiptera	common
<i>Cartodere nodifer</i>	Latridiidae	Coleoptera	common
<i>Cassida vibex</i>	Chrysomelidae	Coleoptera	common
<i>Celastrina argiolus</i>	Lycaenidae	Lepidoptera	common
<i>Cepaea (Cepaea) hortensis</i>	Helicidae	Pulmonata	common
<i>Cepaea (Cepaea) nemoralis</i>	Helicidae	Pulmonata	common
<i>Ceutorhynchus pallidactylus</i>	Curculionidae	Coleoptera	common
<i>Ceutorhynchus pyrrhorhynchus</i>	Curculionidae	Coleoptera	common
<i>Cheilosia albipennis</i>	Syrphidae	Diptera	common
<i>Cheilosia illustrata</i>	Syrphidae	Diptera	common
<i>Chloromyia formosa</i>	Stratiomyidae	Diptera	common
<i>Chorisops tibialis</i>	Stratiomyidae	Diptera	common
<i>Chorthippus brunneus</i>	Acrididae	Orthoptera	common
<i>Chrysolina herbacea</i>	Chrysomelidae	Coleoptera	common
<i>Cicadula quadrinotata</i>	Cicadellidae	Hemiptera	common
<i>Cionus alauda</i>	Curculionidae	Coleoptera	common
<i>Cionus scrophulariae</i>	Curculionidae	Coleoptera	common
<i>Cionus tuberculosus</i>	Curculionidae	Coleoptera	common
<i>Cixius nervosus</i>	Cixiidae	Hemiptera	common
<i>Clausilia (Clausilia) bidentata</i>	Clausiliidae	Pulmonata	common

<i>Cladotomus fulvomaculatus</i>	Miridae	Hemiptera	common
<i>Clubiona pallidula</i>	Clubionidae	Araneae	common
<i>Clubiona phragmitis</i>	Clubionidae	Araneae	common
<i>Clubiona stagnatilis</i>	Clubionidae	Araneae	common
<i>Clusiodes albimanus</i>	Clusiidae	Diptera	common
<i>Coccidula rufa</i>	Coccinellidae	Coleoptera	common
<i>Coccinella septempunctata</i>	Coccinellidae	Coleoptera	common
<i>Coenagrion puella</i>	Coenagrionidae	Odonata	common
<i>Colletes hederae</i>	Colletidae	Hymenoptera	common
<i>Conocephalus fuscus</i>	Conocephalidae	Orthoptera	common
<i>Coreus marginatus</i>	Coreidae	Hemiptera	common
<i>Corixa punctata</i>	Corixidae	Hemiptera	common
<i>Corizus hyoscyami</i>	Rhopalidae	Hemiptera	common
<i>Cornu aspersum</i>	Helicidae	Pulmonata	common
<i>Corticicara gibbosa</i>	Latridiidae	Coleoptera	common
<i>Crepidodera aurata</i>	Chrysomelidae	Coleoptera	common
<i>Crepidodera aurea</i>	Chrysomelidae	Coleoptera	common
<i>Crepidodera fulvicornis</i>	Chrysomelidae	Coleoptera	common
<i>Crudosilis ruficollis</i>	Cantharidae	Coleoptera	common
<i>Cryptoccephalus pusillus</i>	Chrysomelidae	Coleoptera	common
<i>Cymus glandicolor</i>	Lygaeidae	Hemiptera	common
<i>Cyphon coarctatus</i>	Scirtidae	Coleoptera	common
<i>Dasineura pustulans</i>	Cecidomyiidae	Diptera	common
<i>Demetrias atricapillus</i>	Carabidae	Coleoptera	common
<i>Deporaus betulae</i>	Rhynchitidae	Coleoptera	common
<i>Deroceras (Deroceras) reticulatum</i>	Agriolimacidae	Pulmonata	common
<i>Dicranopalpus ramosus</i>	Phalangiidae	Opiliones	common
<i>Dictyla convergens</i>	Tingidae	Hemiptera	local
<i>Dictyna arundinacea</i>	Dictynidae	Araneae	common
<i>Dictyna uncinata</i>	Dictynidae	Araneae	common
<i>Dicyphus (Dicyphus) epilobii</i>	Miridae	Hemiptera	common
<i>Dicyphus (Dicyphus) stachydis</i>	Miridae	Hemiptera	common
<i>Discus (Gonyodiscus) rotundatus</i>	Patulidae	Pulmonata	common
<i>Donacia marginata</i>	Chrysomelidae	Coleoptera	common
<i>Donacia semicuprea</i>	Chrysomelidae	Coleoptera	common
<i>Donacia simplex</i>	Chrysomelidae	Coleoptera	common
<i>Donacia vulgaris</i>	Chrysomelidae	Coleoptera	common
<i>Drupenatus nasturtii</i>	Curculionidae	Coleoptera	[Nb]
<i>Drymus (Sylvadrymus) sylvaticus</i>	Lygaeidae	Hemiptera	common
<i>Dytiscus marginalis</i>	Dytiscidae	Coleoptera	common
<i>Elaphrus cupreus</i>	Carabidae	Coleoptera	common
<i>Eledona agricola</i>	Tenebrionidae	Coleoptera	common
<i>Elmis aenea</i>	Elmidae	Coleoptera	common
<i>Elodes elongata</i>	Scirtidae	Coleoptera	NS
<i>Empis livida</i>	Empididae	Diptera	common

<i>Enallagma cyathigerum</i>	Coenagrionidae	Odonata	common
<i>Enochrus melanocephalus</i>	Hydrophilidae	Coleoptera	common
<i>Enochrus testaceus</i>	Hydrophilidae	Coleoptera	common
<i>Ephemera danica</i>	Ephemeridae	Ephemeroptera	common
<i>Episinus angulatus</i>	Theridiidae	Araneae	common
<i>Epistrophe eligans</i>	Syrphidae	Diptera	common
<i>Episyphus balteatus</i>	Syrphidae	Diptera	common
<i>Epuraea aestiva</i>	Nitidulidae	Coleoptera	common
<i>Epuraea melanocephala</i>	Nitidulidae	Coleoptera	common
<i>Eriothrix rufomaculata</i>	Tachinidae	Diptera	common
<i>Eristalinus sepulchralis</i>	Syrphidae	Diptera	common
<i>Ero cambridgei</i>	Mimetidae	Araneae	common
<i>Erysiphe cruciferarum</i>	Erysiphaceae	Erysiphales	common
<i>Erythromma najas</i>	Coenagrionidae	Odonata	local
<i>Euclidia mi</i>	Erebidae	Lepidoptera	common
<i>Eudonia angustea</i>	Crambidae	Lepidoptera	common
<i>Eupeodes corollae</i>	Syrphidae	Diptera	common
<i>Eurygaster testudinaria</i>	Scutelleridae	Hemiptera	common
<i>Eysarcoris venustissimus</i>	Pentatomidae	Hemiptera	common
<i>Forficula auricularia</i>	Forficulidae	Dermoptera	common
<i>Galba (Galba) truncatula</i>	Lymnaeidae	Hygrophila	common
<i>Galerucella calmariensis</i>	Chrysomelidae	Coleoptera	common
<i>Galerucella nymphaeae</i>	Chrysomelidae	Coleoptera	common
<i>Galerucella tenella</i>	Chrysomelidae	Coleoptera	common
<i>Gallinula chloropus</i>	Rallidae	Gruiformes	common
<i>Gammarus lacustris</i>	Gammaridae	Amphipoda	common
<i>Gastrophysa viridula</i>	Chrysomelidae	Coleoptera	common
<i>Gerris (Gerris) lacustris</i>	Gerridae	Hemiptera	common
<i>Gibbaranea gibbosa</i>	Araneidae	Araneae	common
<i>Gonepteryx rhamni</i>	Pieridae	Lepidoptera	common
<i>Gongylidium rufipes</i>	Limyphiidae	Araneae	common
<i>Grammoptera ruficornis</i>	Cerambycidae	Coleoptera	common
<i>Gymnetron veronicae</i>	Curculionidae	Coleoptera	Nb
<i>Gymnetron villosulum</i>	Curculionidae	Coleoptera	[Nb]
<i>Gymnocheta viridis</i>	Tachinidae	Diptera	common
<i>Gyrinus marinus</i>	Gyrinidae	Coleoptera	common
<i>Gyrinus substriatus</i>	Gyrinidae	Coleoptera	common
<i>Gyrinus urinator</i>	Gyrinidae	Coleoptera	common
<i>Haematopota pluvialis</i>	Tabanidae	Diptera	common
<i>Halesus digitatus</i>	Limnephilidae	Trichoptera	common
<i>Haliplus flavigollis</i>	Haliplidae	Coleoptera	common
<i>Haliplus fluviatilis</i>	Haliplidae	Coleoptera	common
<i>Haliplus lineatocollis</i>	Haliplidae	Coleoptera	common
<i>Harmonia axyridis</i>	Coccinellidae	Coleoptera	common
<i>Harpactea hombergi</i>	Dysderidae	Araneae	common

<i>Helobdella stagnalis</i>	<i>Glossiphoniidae</i>	<i>Rhynchobdellida</i>	common
<i>Helophilus pendulus</i>	<i>Syrphidae</i>	<i>Diptera</i>	common
<i>Helophorus aequalis</i>	<i>Hydrophilidae</i>	<i>Coleoptera</i>	common
<i>Helophorus brevipalpis</i>	<i>Hydrophilidae</i>	<i>Coleoptera</i>	common
<i>Heterogaster urticae</i>	<i>Lygaeidae</i>	<i>Hemiptera</i>	common
<i>Himacerus (Aptus) mirmicoides</i>	<i>Nabidae</i>	<i>Hemiptera</i>	common
<i>Hydrobius fuscipes</i>	<i>Hydrophilidae</i>	<i>Coleoptera</i>	common
<i>Hydroporus palustris</i>	<i>Dytiscidae</i>	<i>Coleoptera</i>	common
<i>Hydroporus planus</i>	<i>Dytiscidae</i>	<i>Coleoptera</i>	common
<i>Hydroporus tessellatus</i>	<i>Dytiscidae</i>	<i>Coleoptera</i>	common
<i>Hygromia (Hygromia) cinctella</i>	<i>Hygromiidae</i>	<i>Pulmonata</i>	common
<i>Hygromoma dimidiata</i>	<i>Staphylinidae</i>	<i>Coleoptera</i>	common
<i>Hygrotus inaequalis</i>	<i>Dytiscidae</i>	<i>Coleoptera</i>	common
<i>Hylyphantes graminicola</i>	<i>Linyphiidae</i>	<i>Araneae</i>	common
<i>Hypomma bituberculatum</i>	<i>Linyphiidae</i>	<i>Araneae</i>	common
<i>Ilybius fenestratus</i>	<i>Dytiscidae</i>	<i>Coleoptera</i>	common
<i>Ilybius fuliginosus</i>	<i>Dytiscidae</i>	<i>Coleoptera</i>	common
<i>Ischnodemus sabuleti</i>	<i>Lygaeidae</i>	<i>Hemiptera</i>	common
<i>Ischnura elegans</i>	<i>Coenagrionidae</i>	<i>Odonata</i>	common
<i>Isoperla grammatica</i>	<i>Perlodidae</i>	<i>Plecoptera</i>	common
<i>Iteomyia major</i>	<i>Cecidomyiidae</i>	<i>Diptera</i>	common
<i>Javesella obscurella</i>	<i>Delphacidae</i>	<i>Hemiptera</i>	common
<i>Kaestneria pullata</i>	<i>Linyphiidae</i>	<i>Araneae</i>	common
<i>Kateretes pusillus</i>	<i>Kateretidae</i>	<i>Coleoptera</i>	common
<i>Laccobius bipunctatus</i>	<i>Hydrophilidae</i>	<i>Coleoptera</i>	common
<i>Laccobius minutus</i>	<i>Hydrophilidae</i>	<i>Coleoptera</i>	common
<i>Laccophilus minutus</i>	<i>Dytiscidae</i>	<i>Coleoptera</i>	common
<i>Laetiporus sulphureus</i>	<i>Fomitopsidaceae</i>	<i>Polyporales</i>	common
<i>Larinoides cornutus</i>	<i>Araneidae</i>	<i>Araneae</i>	common
<i>Lasius niger</i>	<i>Formicidae</i>	<i>Hymenoptera</i>	common
<i>Lasius platythorax</i>	<i>Formicidae</i>	<i>Hymenoptera</i>	common
<i>Lathys humilis</i>	<i>Dictynidae</i>	<i>Araneae</i>	common
<i>Leiobunum rotundum</i>	<i>Phalangiidae</i>	<i>Opiliones</i>	common
<i>Lejogaster metallina</i>	<i>Syrphidae</i>	<i>Diptera</i>	local
<i>Leptopterna dolabrata</i>	<i>Miridae</i>	<i>Hemiptera</i>	common
<i>Libellula depressa</i>	<i>Libellulidae</i>	<i>Odonata</i>	common
<i>Limacus flavus</i>	<i>Limacidae</i>	<i>Pulmonata</i>	common
<i>Limax maximus</i>	<i>Limacidae</i>	<i>Pulmonata</i>	common
<i>Limnia paludicola</i>	<i>Sciomyzidae</i>	<i>Diptera</i>	common
<i>Linyphia triangularis</i>	<i>Linyphiidae</i>	<i>Araneae</i>	common
<i>Liocoris tripustulatus</i>	<i>Miridae</i>	<i>Hemiptera</i>	common
<i>Lochmaea caprea</i>	<i>Chrysomelidae</i>	<i>Coleoptera</i>	common
<i>Lochmaea crataegi</i>	<i>Chrysomelidae</i>	<i>Coleoptera</i>	common
<i>Longitarsus dorsalis</i>	<i>Chrysomelidae</i>	<i>Coleoptera</i>	common
<i>Longitarsus rubiginosus</i>	<i>Chrysomelidae</i>	<i>Coleoptera</i>	common

<i>Lucilia sericata</i>	<i>Calliphoridae</i>	<i>Diptera</i>	common
<i>Lymnaea stagnalis</i>	<i>Lymnaeidae</i>	<i>Hygrophila</i>	common
<i>Macrophya annulata</i>	<i>Tenthredinidae</i>	<i>Hymenoptera</i>	common
<i>Macrophya ribis</i>	<i>Tenthredinidae</i>	<i>Hymenoptera</i>	common
<i>Malachius bipustulatus</i>	<i>Malachiidae</i>	<i>Coleoptera</i>	common
<i>Maniola jurtina</i>	<i>Nymphalidae</i>	<i>Lepidoptera</i>	common
<i>Megamelodes quadrimaculatus</i>	<i>Delphacidae</i>	<i>Hemiptera</i>	local
<i>Meligethes aeneus</i>	<i>Nitidulidae</i>	<i>Coleoptera</i>	common
<i>Merodon equestris</i>	<i>Syrphidae</i>	<i>Diptera</i>	common
<i>Metellina mengei</i>	<i>Tetragnathidae</i>	<i>Araneae</i>	common
<i>Metellina segmentata</i>	<i>Tetragnathidae</i>	<i>Araneae</i>	common
<i>Micaria pulicaria</i>	<i>Gnaphosidae</i>	<i>Araneae</i>	common
<i>Microlinyphia impigra</i>	<i>Linyphiidae</i>	<i>Araneae</i>	common
<i>Micromus variegatus</i>	<i>Hemerobiidae</i>	<i>Neuroptera</i>	common
<i>Microvelia (Microvelia) reticulata</i>	<i>Veliidae</i>	<i>Hemiptera</i>	common
<i>Misumena vatia</i>	<i>Thomisidae</i>	<i>Araneae</i>	common
<i>Monacha (Monacha) cantiana</i>	<i>Hygromiidae</i>	<i>Pulmonata</i>	common
<i>Myathropa florea</i>	<i>Syrphidae</i>	<i>Diptera</i>	common
<i>Myrmica ruginodis</i>	<i>Formicidae</i>	<i>Hymenoptera</i>	common
<i>Mystacides longicornis</i>	<i>Leptoceridae</i>	<i>Trichoptera</i>	common
<i>Nebrioporus elegans</i>	<i>Dytiscidae</i>	<i>Coleoptera</i>	common
<i>Nedyus quadrimaculatus</i>	<i>Curculionidae</i>	<i>Coleoptera</i>	common
<i>Nemotelus pantherinus</i>	<i>Stratiomyidae</i>	<i>Diptera</i>	common
<i>Nemoura cinerea</i>	<i>Nemouridae</i>	<i>Plecoptera</i>	common
<i>Nemurella pictetii</i>	<i>Nemouridae</i>	<i>Plecoptera</i>	common
<i>Neoascia geniculata</i>	<i>Syrphidae</i>	<i>Diptera</i>	common
<i>Neoascia tenur</i>	<i>Syrphidae</i>	<i>Diptera</i>	common
<i>Neocoenorrhinus aequatus</i>	<i>Rhynchitidae</i>	<i>Coleoptera</i>	common
<i>Noterus clavicornis</i>	<i>Noteridae</i>	<i>Coleoptera</i>	common
<i>Notonecta glauca</i>	<i>Notonectidae</i>	<i>Hemiptera</i>	common
<i>Notonecta viridis</i>	<i>Notonectidae</i>	<i>Hemiptera</i>	common
<i>Nyctia halterata</i>	<i>Sarcophagidae</i>	<i>Diptera</i>	common
<i>Ochina ptinoides</i>	<i>Anobiidae</i>	<i>Coleoptera</i>	common
<i>Ochliodes sylvanus</i>	<i>Hesperiidae</i>	<i>Lepidoptera</i>	common
<i>Ochthebius minimus</i>	<i>Hydraenidae</i>	<i>Coleoptera</i>	common
<i>Oedemera lutea</i>	<i>Oedemeridae</i>	<i>Coleoptera</i>	common
<i>Oedemera nobilis</i>	<i>Oedemeridae</i>	<i>Coleoptera</i>	common
<i>Oligia fasciuncula</i>	<i>Noctuidae</i>	<i>Lepidoptera</i>	common
<i>Opilio parietinus</i>	<i>Phalangiidae</i>	<i>Opiliones</i>	common
<i>Opilio parietinus</i>	<i>Phalangiidae</i>	<i>Opiliones</i>	common
<i>Orchesella cincta</i>	<i>Entomobryidae</i>	<i>Collembola</i>	common
<i>Orius (Heterorius) laticollis</i>	<i>Anthocoridae</i>	<i>Hemiptera</i>	common
<i>Orthops (Orthops) campestris</i>	<i>Miridae</i>	<i>Hemiptera</i>	common
<i>Oxyloma (Oxyloma) elegans</i>	<i>Succineidae</i>	<i>Pulmonata</i>	common
<i>Pachygaster atra</i>	<i>Stratiomyidae</i>	<i>Diptera</i>	common

<i>Pachynatha clercki</i>	Tetragnathidae	Araneae	common
<i>Paederus littoralis</i>	Staphylinidae	Coleoptera	common
<i>Paederus riparius</i>	Staphylinidae	Coleoptera	common
<i>Paidiscura pallens</i>	Theridiidae	Araneae	common
<i>Palomena prasina</i>	Pentatomidae	Hemiptera	common
<i>Panorpa germanica</i>	Panorpidae	Mecoptera	common
<i>Pantilius (Pantilius) tunicatus</i>	Miridae	Hemiptera	local
<i>Paracorymbia fulva</i>	Cerambycidae	Coleoptera	RDB 3
<i>Paradromius linearis</i>	Carabidae	Coleoptera	common
<i>Pararge aegeria</i>	Nymphalidae	Lepidoptera	common
<i>Pardosa amentata</i>	Lycosidae	Araneae	common
<i>Pardosa saltans</i>	Lycosidae	Araneae	common
<i>Pegomya solennis</i>	Anthomyiidae	Diptera	common
<i>Peponocranium ludicum</i>	Linyphiidae	Araneae	common
<i>Phaedon armoraciae</i>	Chrysomelidae	Coleoptera	common
<i>Phaedon cochleariae</i>	Chrysomelidae	Coleoptera	common
<i>Pherbellia schoenherri</i>	Sciomyzidae	Diptera	common
<i>Philaenus spumarius</i>	Aphrophoridae	Hemiptera	common
<i>Philodromus dispar</i>	Philodromidae	Araneae	common
<i>Pholcomma gibbum</i>	Theridiidae	Araneae	common
<i>Pholidoptera griseoaptera</i>	Tettigoniidae	Orthoptera	common
<i>Phratora vulgatissima</i>	Chrysomelidae	Coleoptera	common
<i>Phyllobius pyri</i>	Curculionidae	Coleoptera	common
<i>Phyllobius roboretanus</i>	Curculionidae	Coleoptera	common
<i>Phyllobius virideae</i>	Curculionidae	Coleoptera	common
<i>Phylloneta sisypheia</i>	Theridiidae	Araneae	common
<i>Phyllopertha horticola</i>	Rutelidae	Coleoptera	common
<i>Physa fontinalis</i>	Physidae	Hygrophila	common
<i>Phytomyza ilicis</i>	Agromyzidae	Diptera	common
<i>Pieris brassicae</i>	Pieridae	Lepidoptera	common
<i>Pieris rapae</i>	Pieridae	Lepidoptera	common
<i>Pinalitus cervinus</i>	Miridae	Hemiptera	common
<i>Pirata piraticus</i>	Lycosidae	Araneae	common
<i>Pisaura mirabilis</i>	Pisauridae	Araneae	common
<i>Piscicola geometra</i>	Piscicolidae	Rhynchobdellida	common
<i>Planorbis carinatus</i>	Planorbidae	Hygrophila	common
<i>Planorbis planorbis</i>	Planorbidae	Hygrophila	common
<i>Plateumaris rustica</i>	Chrysomelidae	Coleoptera	NS
<i>Platycheirus albimanus</i>	Syrphidae	Diptera	common
<i>Platycheirus rosanum</i>	Syrphidae	Diptera	common
<i>Platystethus arenarius</i>	Staphylinidae	Coleoptera	common
<i>Podops inuncta</i>	Pentatomidae	Hemiptera	common
<i>Pogonocherus hispidus</i>	Cerambycidae	Coleoptera	common
<i>Poophagus sisymbrii</i>	Curculionidae	Coleoptera	common
<i>Prasocuris junci</i>	Chrysomelidae	Coleoptera	common

<i>Propyles quattuordecimpunctata</i>	Coccinellidae	Coleoptera	common
<i>Psammoecus bipunctatus</i>	Silvanidae	Coleoptera	common
<i>Psyche casta</i>	Psychidae	Lepidoptera	common
<i>Psylla alni sensu stricto</i>	Psyllidae	Hemiptera	common
<i>Psylliodes affinis</i>	Chrysomelidae	Coleoptera	common
<i>Psylliodes chrysocephala</i>	Chrysomelidae	Coleoptera	common
<i>Psylliodes dulcamarae</i>	Chrysomelidae	Coleoptera	common
<i>Ptilinus pectinicornis</i>	Anobiidae	Coleoptera	common
<i>Ptychoptera contaminata</i>	Ptychopteridae	Diptera	common
<i>Pyrausta aurata</i>	Crambidae	Lepidoptera	common
<i>Pyrrhosoma nymphula</i>	Coenagrionidae	Odonata	common
<i>Radix balthica</i>	Lymnaeidae	Hygrophila	common
<i>Rhagomycha fulva</i>	Cantharidae	Coleoptera	common
<i>Rhagomycha lignosa</i>	Cantharidae	Coleoptera	common
<i>Rhinoncus pericarpinus</i>	Curculionidae	Coleoptera	common
<i>Rhizomnium punctatum</i>	Cinclidiaceae	Bryales	common
<i>Rhogogaster viridis</i>	Tenthredinidae	Hymenoptera	common
<i>Rhyzobius litura</i>	Coccinellidae	Coleoptera	common
<i>Robertus lividus</i>	Theridiidae	Araneae	common
<i>Rutpela maculata</i>	Cerambycidae	Coleoptera	common
<i>Salpingus planirostris</i>	Salpingidae	Coleoptera	common
<i>Salticus scenicus</i>	Salticidae	Araneae	common
<i>Scathophaga stercoraria</i>	Scathophagidae	Diptera	common
<i>Scolopostethus thomsoni</i>	Lygaeidae	Hemiptera	common
<i>Sepedophilus littoreus</i>	Staphylinidae	Coleoptera	common
<i>Sesia bembeciformis</i>	Sesiidae	Lepidoptera	common
<i>Sialis lutaria</i>	Sialidae	Megaloptera	common
<i>Sicus ferrugineus</i>	Conopidae	Diptera	common
<i>Sigara (Subsigara) fossarum</i>	Corixidae	Hemiptera	common
<i>Sisyra nigra</i>	Sisyridae	Neuroptera	common
<i>Sitona lineatus</i>	Curculionidae	Coleoptera	common
<i>Sphaeridium scarabaeoides</i>	Hydrophilidae	Coleoptera	common
<i>Stenus canescens</i>	Staphylinidae	Coleoptera	Nb
<i>Stenus latifrons</i>	Staphylinidae	Coleoptera	common
<i>Stigmella aurella</i>	Nepticulidae	Lepidoptera	common
<i>Stratiomys potamida</i>	Stratiomyidae	Diptera	common
<i>Succinea putris</i>	Succineidae	Pulmonata	common
<i>Synanthedon bembeciformis</i>	Sesiidae	Lepidoptera	common
<i>Tachina fera</i>	Tachinidae	Diptera	common
<i>Tachyporus chrysomelinus</i>	Staphylinidae	Coleoptera	common
<i>Telmatophilus caricis</i>	Cryptophagidae	Coleoptera	common
<i>Telmatophilus typhae</i>	Cryptophagidae	Coleoptera	common
<i>Tetanocera arrogans</i>	Sciomyzidae	Diptera	common
<i>Tetanocera ferruginea</i>	Sciomyzidae	Diptera	common

<i>Tetragnatha extensa</i>	Tetragnathidae	Araneae	common
<i>Tetragnatha montana</i>	Tetragnathidae	Araneae	common
<i>Theridiosoma gemmosum</i>	Theridiosomatidae	Araneae	NS
<i>Thryogenes nereis</i>	Eriihiniidae	Coleoptera	common
<i>Tibellus oblongus</i>	Philodromidae	Araneae	common
<i>Trioza urticae</i>	Trioziidae	Hemiptera	common
<i>Tytthaspis sedecimpunctata</i>	Coccinellidae	Coleoptera	common
<i>Valvata piscinalis</i>	Valvatidae	Pulmonata	common
<i>Velia caprai</i>	Veliidae	Hemiptera	common
<i>Vespa vulgaris</i>	Vespidae	Hymenoptera	common
<i>Volucella bombylans</i>	Syrphidae	Diptera	common

Appendix 2. Status categories for rare and Notable species

Red Data Book Category 1 (RDB 1) – Endangered

Definition.

Taxa in danger of extinction in Great Britain and whose survival is unlikely if the causal factors continue operating.

Included are those taxa whose numbers have been reduced to a critical level or whose habitats have been so dramatically reduced that they are deemed to be in immediate danger of extinction. Also included are some taxa that are possibly extinct.

Criteria.

Species which are known or believed to occur as only a single population within one 10 km square of the National Grid.

Species which only occur in habitats known to be especially vulnerable.

Species which have shown a rapid or continuous decline over the last twenty years and are now estimated to exist in five or fewer 10 km squares.

Species which are possibly extinct but have been recorded this century and if rediscovered would need protection.

Red Data Book Category 2 (RDB 2) - Vulnerable

Definition.

Taxa believed likely to move into the endangered category in the near future if the causal factors continue operating.

Included are taxa of which most or all of the populations are decreasing because of over-exploitation, extensive destruction of habitat or other environmental disturbance; taxa with populations that have been seriously depleted and whose ultimate security is not yet

assured; and taxa with populations that are still abundant but are under threat from serious adverse factors throughout their range.

Criteria.

Species declining throughout their range.

Species in vulnerable habitats.

Red Data Book Category 3 (RDB 3) – Rare

Definition.

Taxa with small populations in Great Britain that are not at present endangered or vulnerable, but are at risk.

These taxa are usually localised within restricted geographical areas or habitats or are thinly scattered over a more extensive range.

Criterion.

Species which are estimated to exist in only fifteen or fewer 10 km squares. *This criterion may be relaxed where populations are likely to exist in over fifteen 10 km squares but occupy small areas of especially vulnerable habitat*

Nationally Scarce Category A - Notable A (Na)

Definition.

Taxa which do not fall within RDB categories but which are none-the-less uncommon in Great Britain and are thought to occur in 30 or fewer 10 km squares of the National Grid or, for less well recorded groups, within seven or fewer vice-counties.

Nationally Scarce Category B - Notable B (Nb)

Definition.

Taxa which do not fall within RDB categories but which are none-the-less uncommon in Great Britain and are thought to occur in between 31 and 100 10 km squares of the National Grid or, for less well recorded groups, within eight and twenty vice-counties.

Nationally Scarce - Notable (N)

Definition.

Taxa which do not fall within RDB categories but which are none-the-less uncommon in Great Britain and are thought to occur in between 16 to 100 10 km squares of the National Grid. Species within this category are often too poorly known for their status to be more precisely estimated.

Summary of the IUCN categories and criteria.

GB Rarity Status categories and criteria

- **Nationally Rare (NR)**

Native species which have not been recorded from more than 15 British hectads since 31st December 1979 and where there is reasonable confidence that exhaustive recording would not find them in more than 15 hectads. This category includes species which are probably extinct.

- **Nationally Scarce (NS)**

Native species which are not regarded as Nationally Rare AND which have not been recorded from more than 100 British hectads since 31st December 1979 and where there is reasonable confidence that exhaustive recording would not find them in more than 100 hectads.

Other species status terminology.

- **Local.** Species that are restricted in distribution either geographically or by habitat. Also used for species that are widespread but infrequently encountered, e.g. encountered in no more than 300 10km squares of the national Ordnance Survey grid since 1970. Or those species listed as such, based upon modern geographical data, by ISIS (2010) and/or relevant recording schemes.
- **Common.** Generally widespread throughout the UK.

MILL LAKE, ANDOVER

Water Assessment and Management



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This report concerns Mill Lake, which forms part of the Rooksbury Mill, Local Nature Reserve, Andover, Grid Reference SU35599 44607. The lake is approximately 2.1ha (5acres), with an inlet on the north bank from the River Anton and outlet on the south west side of the lake. The lake has a mixed coarse fish population but is lightly stocked, with a limited amount of angling permitted. The lake is overwhelmed with fibrous blanket weed, some of which has been removed using a weed cutting boat, but the water surface has large areas of the nuisance algae. Barley straw has also been used to control the blanket weed growth, but the volume of weed has exceeded the capacity of the straw to affect it.

Water analysis

The water chemistry on the lake was assessed using electronic meters for dissolved oxygen, conductivity, total dissolved solids, pH and temperature, a Palintest 7100 photometer was used for chemical assay the results are given in Table 1.

Table 1. Water Analysis, Mill Lake

	Inlet	Outlet
Colour	Clear	Clear
pH	7.1	7.06
Total Ammonia (mg per litre as N)	0.01	0.05
Ammonia as NH ³ (mg per litre)	0.01	0.06
Ammonia as NH ⁴ (mg per litre)	0.01	0.06
Nitrite (mg per litre as N)	0.046	0.058
Nitrite as NO ₂ (mg per litre)	0.151	0.191
Nitrate (mg per litre as N)	10.38	8.76
Nitrate as NO ₃ (mg per litre)	45.98	38.8
Phosphate (mg per litre as P)	0.15	0.08
Phosphate as PO ₄ (mg per litre)	0.44	0.24
Total Alkalinity (as calcium carbonate mg per litre)	277	286
Total Alkalinity HCO ₃ ⁻	338	349
Total Alkalinity CO ₃ ²⁻	166	172
Redox (mV)	197	103
Total Dissolved Solids (g per litre)	0.26	0.26
Conductivity (mS)	0.52	0.52

Comments

The main area of concern is the nutrient content of the lake, comprising the nitrogen (nitrite and nitrate) and phosphate content which are causing the water to become eutrophic (nutrient enriched). Using a Trophic State Index, the water in Mill Lake falls

into the category of poor based on the nitrogen and phosphorous content. It is the concentration of these nutrients in the water which are promoting the growth of the nuisance *Cladophora* sp. (blanket weed).

In addition to the *Cladophora*, the dominant phytoplankton was the diatom *Syndra* sp., with small numbers of the Cyanobacteria (blue-green algae), *Lyngbya* sp. and some motile green algae, *Chlamydomonas* sp. The *Lyngbya* sp. was present in very low numbers, less than 10 per ml of water, which does not represent a health hazard, numbers in excess of 20,000 per ml are regarded as serious. I should also add that it is possible to find representative species of Cyanobacteria in most lakes in England.

Cladophora thrives in eutrophic waters, forming the extensive mats which are plaguing Mill Lake, which as they decay cause the water to become anoxic, affecting all freshwater life. This nuisance algae responds rapidly to changes in day length and as soon as the day length noticeably increases in late January, early February, it will start growing again. Although cutting the *Cladophora* is the best current means of control, the algae will grow from any fragments left in the water, which potentially causes the weed to proliferate.

Recommendations

1) Barley straw – In the presence of oxygen, the microbial decomposition of barley straw releases hydrogen peroxide, termed 'algae inhibiting factor', kills the algae. The application of barley straw is roughly 50g per square metre of water surface, ideally loosely packing the barley straw into Christmas tree netting or rafts (Figure 1). At temperatures of 10°C and below the barley straw will take about 6 – 8 weeks to release the algae inhibiting factor, at temperatures approaching 20°C the barley straw becomes effective within 2 – 3 weeks, with inhibition of algae lasting about 4 – 6 weeks depending on temperature. Ideally, the barley straw needs to be placed on the lake in late winter, January to February to inhibit the growth of the blanket weed. The best use of barley straw is to create a rolling programmes so that new batches of barley straw are added to the lake before the existing barley straw ceases production of the inhibiting factor. Always remove the expired barley straw as this also becomes subject to decomposition, becoming a further source of nitrogen in the water.

It may be worth creating a single line of barley straw sausages, crossing the lake from shore to shore rather than using circles and possibly rather than a line of sausages a couple of rafts at the inlet.

Rather than using plastic bottles, which are a little unsightly it may be better to using fishing net floats, which can be tied to the barley straw sausages or rafts. The following can supply the floats and the price to give you an idea, as you can see there is variation in price, associated with the weight of the float but worth an internet search:

www.collins.co.uk floats 0.36p each

www.advancednetting.co.uk 0.24p each

www.gaelforcemarine.co.uk 0.64p each

www.coastalnets.co.uk £6.50 per 10



Figure 1. Barley straw raft

2) Planting – Planting is the key to controlling nuisance algae as this is the best means of removing nutrients from the lake. It would be worth planting reeds, possibly common reed (*Phragmites australis*) on either side of the inlet of the River Anton into the lake. The reeds are extremely good at extracting nutrients and any potential pollutants from the water, as it enters the lake. Common reed can be very invasive and needs to be kept under control, I would certainly advocate cutting back the dead stems and flower

heads annually in the autumn. Planting the margins will also help to compete with the algae for available nutrients, fool's watercress *Apium nodiflorum* is a good marginal plant, grows early in the year, competing with the algae and produces white flowers for the insect life. There are already marginal plants on the lake and cuttings of these could be transplanted around the lake shores, this is best undertaken in the spring as the water is warming and allows the plants the summer months to become fully established.

3) Dead heading – All dead plant leaves, flowers and stems which drop into the water also add to the nitrogen and phosphate concentration, which dead heading can reduce but I'm not sure how much impact that might have on terrestrial insect life.

4) Siltex® - Siltex® is a champagne chalk and is excellent for controlling nuisance *Cladophora*, breaking down the organic deposits in the lake and effectively reducing the available nutrients. The use of Siltex® will often promote the growth of the macrophytes on the lake. The application rate is 1 tonne per 0.4ha (1 acre), usually it is applied off a boat, to allow the propeller to mix it into the water, Kingcombe Aquacare would be able to undertake the work. The application of Siltex® may be piecemeal over a few years and it may be that treating just part of the lake adjacent to the inlet will have a significant impact on the growth of the *Cladophora*. Treatment is usually undertaken in the autumn or winter, as there is the possibility that as the Siltex® breaks down the organic material it can impact on the dissolved oxygen. The autumn winter are better as oxygen is more soluble in cold water. As a guide, one company charge £252 per tonne of Siltex® including delivery, however Kingcombe Aquacare may have a different source of champagne chalk other than the branded product.

5) Phoslock® - Phoslock® is an extremely effective product for stripping the water of phosphorous and controlling nuisance algae. The phosphorous is chemically locked into the sediment and therefore the algae are unable to grow, macrophytes are unaffected because the root system takes the nutrients from the sediment. It has been successfully used for natural swimming lakes and for treating the Serpentine in 2012 for the triathlon in the Olympic Games. While it sounds like the dream product, it is astronomically expensive.

5) Combination approach – although I have identified several methods of controlling the proliferation of *Cladophora* the best method is a combination. Even if you opt for

using Siltex®, it would be worth combining this with the use of barley straw, especially if the Siltex® is used to treat the lake piecemeal.

Finally, there is no such thing as a quick fix when it comes to the aquatic environment and whatever remedial action is undertaken, it may take several years before the best result is achieved, it may take a few years to finally achieve eradication of the *Cladophora*.

APPENDIX III

Parks & Countryside Event Guide 2026



Anton Lakes LNR, Andover

Grid reference: SU357466

Anton Lakes were formed as a result of gravel extraction. The lake outflow is the headwaters of the River Anton. The springs, lake, river, and meadows around the edge of the site provide a

mosaic of habitats, including chalk grassland, watercress beds, and water meadows. Cattle now graze throughout the summer months and help to support the diverse range of wildflowers. Many birds can also be found on the lakes and streams including kingfishers and water rail.

Ladies Walk, Andover

Grid reference: SU369445

Located on the south eastern edge of Andover, Ladies Walk comprises of three moderately steep north facing meadows. The chalk grassland on site supports a diverse array of butterflies and other invertebrates, as well as vertebrates such as common lizards and slow worms. The meadows are grazed throughout the summer to improve their sward structure. The Ladies Walk, which borders the site, dates back to 1785.

Rooksbury Mill LNR, Andover

Grid reference: SU356443

Rooksbury Mill, also formed through gravel extraction and once a trout fishery, now plays host to a variety of wildlife including otters, water voles, and kingfishers. Its mosaic of chalk grassland and riparian habitats provide a rich diversity of wildlife.

Harewood Common, Andover

Grid reference: SU357466

Comprising of nearly 10 hectares of chalk grassland, Harewood Common is rich in wildlife and provides a refreshing contrast to the nearby Urban Park. Originally an agricultural field, the common backs onto the ancient woodland of Harewood Forest and is managed through grazing during the summer months. This creates an ideal habitat for insects, birds, and bats to thrive.

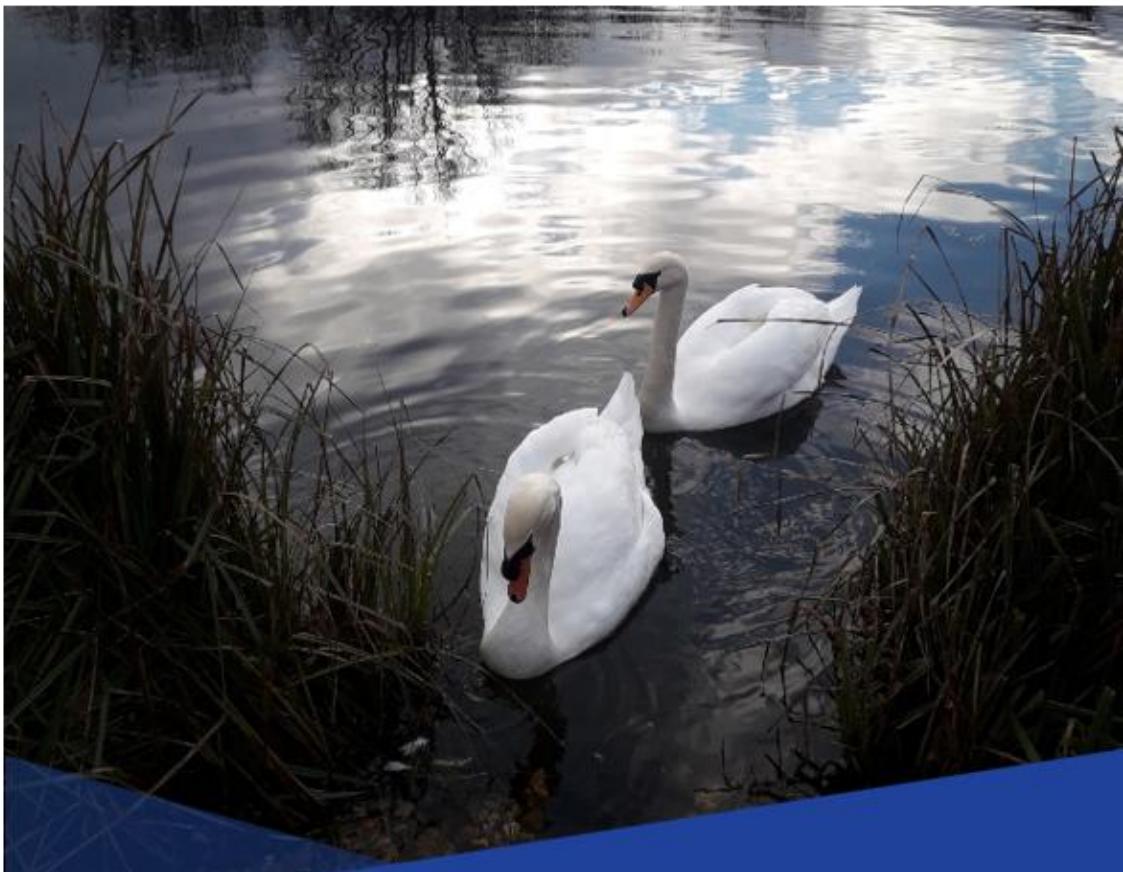
Ox Drove Meadow, Andover

Grid reference: SU357466

Comprising of just over three hectares of chalk grassland, Ox Drove Meadow is made up of a gently sloping path which encompasses the site. New trees planted around the site are helping to establish a robust hedgerow around its boundary which is important for the sites' inhabitants which include a wide variety of birds and insects, as well as the Hazel Dormouse which makes the site its home.

Parks and Countryside

Events and Activities 2026



Test Valley Parks and Countryside Team

The team work to maintain and develop the borough's green spaces which include countryside sites, Nature Reserves, parks, and cemeteries.

Please contact the countryside officers in advance for more details.

Further information is available on www.testvalley.gov.uk

Or contact the Countryside Officers on 01264 368000, or email community&leisure@testvalley.gov.uk

Local Conservation Groups

The Anton River Conservation Association (TARCA)

Thank you to everyone who has volunteered throughout 2025 to help maintain and enhance our green spaces.

New volunteers, including families, are always welcome to help with the on-going programme of events.

North Test Valley

 TestValleyBC

 TestValleyBC

www.testvalley.gov.uk

Test Valley
Borough Council


T.A.R.C.A.
THE ANTON RIVER CONSERVATION ASSOCIATION

Parks and Countryside Events and Activities Guide 2026

North Test Valley

Date	Time	Task	Venue	Meeting Place (what3words)
Thur 8th Jan	10am- 2pm	General Site Maintenance	Bury Hill Meadows	Car park: //loser.waddle.pilots
Sun 11th Jan	10am- 2pm	Wildlife Corridor Clean-Up	Ox Drove Meadows	Picket Piece sports ground: ///guitar.thighs.jubilant
Sat 24 th Jan	10am- 2pm	Pathway coppicing	Harewood Common	Car park by P20 Co-op: ///backdrop.screamed.shallower
Thur 5th Feb	10am- 2pm	Hedgerow Rejuvenation	Ladies Walk LNR	Hedge End Road playing field: //gain.loft.fire
Sun 8th Feb	10am- 2pm	Coppicing	Anton Lakes LNR	Car park: ///warthog.code.starch
Sun 22nd Feb	10am- 2pm	Watercress bed Maintenance	Anton Lakes LNR	Car park: ///warthog.code.starch
Thur 5th March	10am- 2pm	Woody Debris Mattress Creation	Anton Lakes LNR	Car park: ///warthog.code.starch
Sat 28th March	10am- 2pm	General Site Maintenance	Rooksbury Mill LNR	Car park: ///town.rider.rank
Sun 12th April	10am- 2pm	Big Litter Pick	Anton Lakes LNR	Car park: ///warthog.code.starch
Mon 4th May	5am onwards	Dawn chorus	Rooksbury Mill LNR	Car park: ///town.rider.rank
Sun 17th May	10am- 2pm	Big Litter Pick	Town Centre	Car park rear of ASDA: ///canny.movies.hunt
Sat 13th June	10am- 2pm	Non-native Species Removal	Rooksbury Mill LNR	Car park: ///town.rider.rank
Sun 12th July	10am- 2pm	Butterfly Discovery Day	Bury Hill Meadows	Car park: //loser.waddle.pilots
Sat 1st Aug	10am- 2pm	Discovery Day	Anton Lakes LNR	Car park: ///warthog.code.starch
Sun 16th Aug	10am- 2pm	Summer walk	East Anton to Smannell	Sports ground car park: ///tearfully.dozens.sporting
Sun 23rd Aug	10am- 2pm	Non-native species removal	Rooksbury Mill LNR	Car park: ///town.rider.rank
Fri 28th Aug	Pre-booked only	Bat Walk	Anton Lakes LNR	Car park: ///warthog.code.starch
Sat 5th Sept	10am- 2pm	General Site Maintenance	Harewood Common	Car park by P20 Co-op: ///backdrop.screamed.shallower
Fri 11th Sept	Pre-booked only	Bat Walk	Rooksbury Mill LNR	Car park: ///town.rider.rank
Sun 20th Sept	10am- 2pm	Autumn Tasks	Rooksbury Mill LNR	Car park: ///town.rider.rank
Thur 1st Oct	10am- 2pm	Scrub Removal	Ladies Walk LNR	Hedge End Road playing field: //gain.loft.fire
Sat 10th Oct	10am- 2pm	Scrub Removal	Ladies Walk LNR	Hedge End Road playing field: //gain.loft.fire
Sun 25th Oct	10am- 2pm	ASDA riverbank cutting	Behind ASDA	Car park rear of ASDA: ///canny.movies.hunt
Thur 5th Nov	10am- 2pm	Coppicing	Harewood Common	Car park by P20 Co-op: ///backdrop.screamed.shallower
Sat 14th Nov	10am- 2pm	Coppicing and woody debris mattress creation	Anton Lakes LNR	Car park: ///warthog.code.starch
Sun 29th Nov	10am- 2pm	Winter Tasks	Rooksbury Mill LNR	Car park: ///town.rider.rank
Thur 3rd Dec	10am- 2pm	Winter Tasks	Rooksbury Mill LNR	Car park: ///town.rider.rank
Sat 12th Dec	10am- 2pm	Winter Tasks	Rooksbury Mill LNR	Car park: ///town.rider.rank

APPENDIX IV

EDUCATIONAL VISITOR GUIDE



Rooksbury Mill Local Nature Reserve Educational Visitor Guide and Activity Sheets

Habitats Landscapes

Plant and Animal Lifecycles

Copies can be viewed at :

<https://www.testvalley.gov.uk/communityandleisure/naturereserves/free-activity-visit-schools-test-valley-nature-res>

Some comments from the Schools Nature Reserve Visit 2019

“The children really enjoyed the visit. Thank you for making it so enjoyable for them. We did some work on pond dipping and understanding of the journey of a river before our visit. We plan to follow up with work around habitats on our return to school.”

“Before our visit we did some work on animal classification and will continue this upon our return.”

“A fantastic half day with patient and engaging leaders. It is great to visit sites in the local area that the children may visit with their family and give them a better insight into the nature and care need to support these places. Thank you for organising these events.”